A new molecular assay on barley seed to quantify Ustilago nuda and predict field infection levels







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Date: 01.06.2025



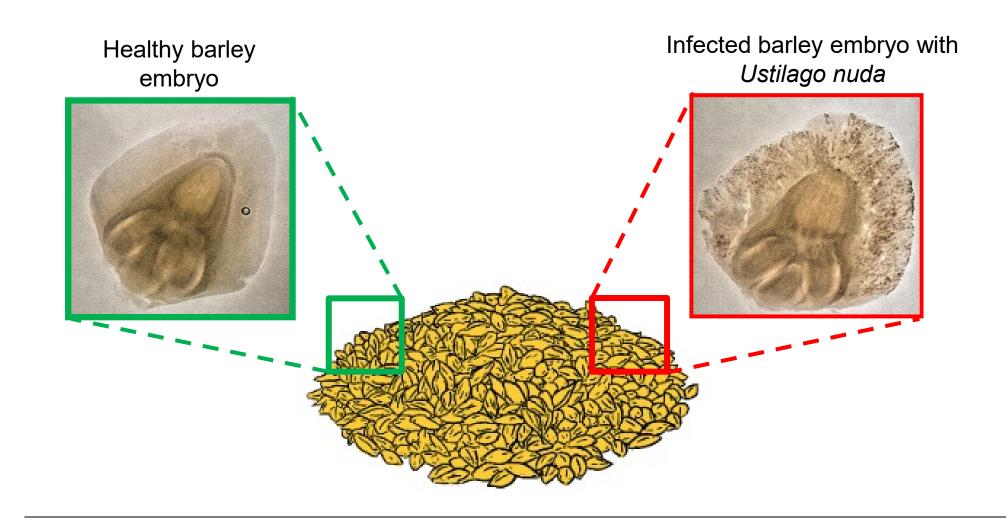


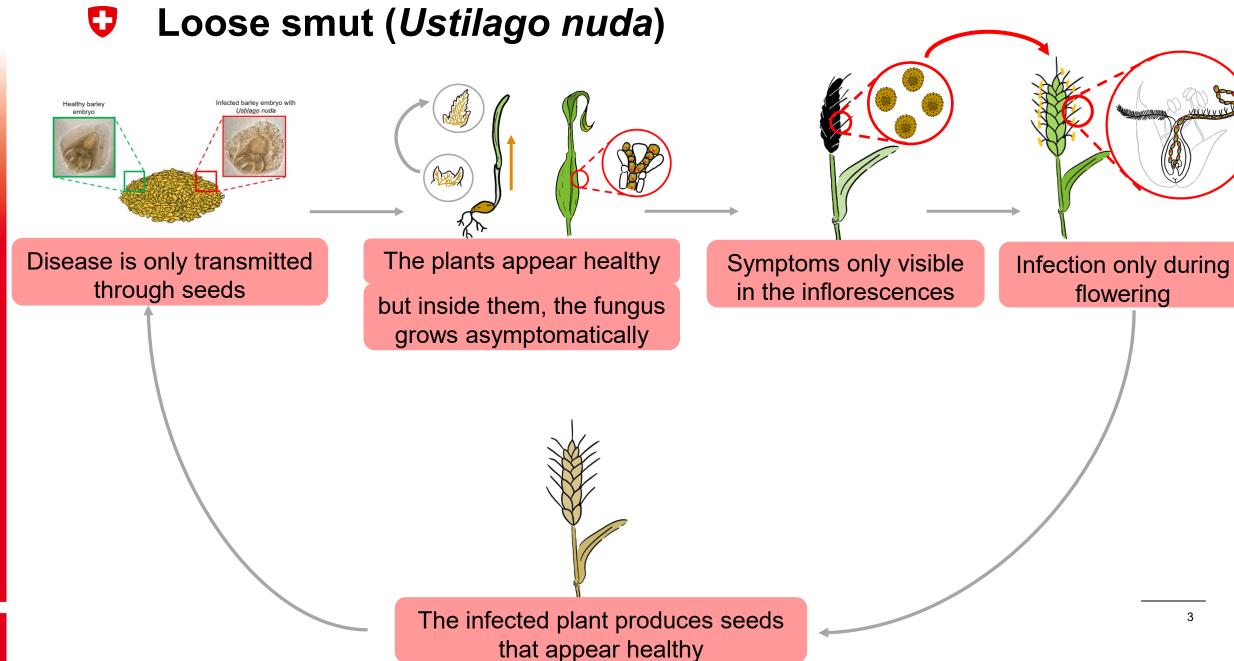






Ustilago nuda)

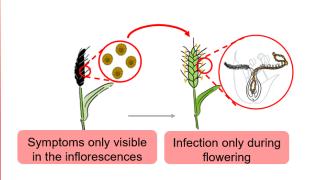


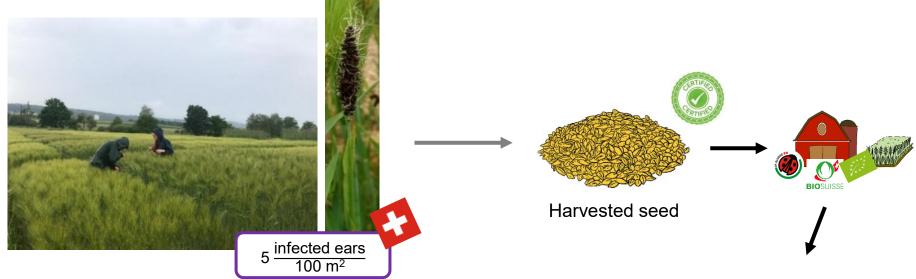


O

Certified seed and smut infection











Unexpectedly high infection rate

Field monitoring problems



Time-consuming and laborious

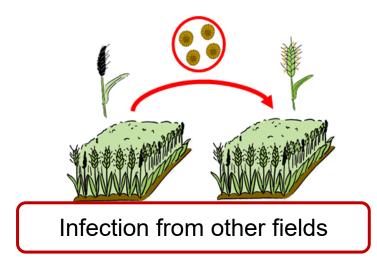


Infections are difficult to detect



Sensitive to environmental influences

Infected ears are less visible because the spores are washed away by rain or blown away by the wind

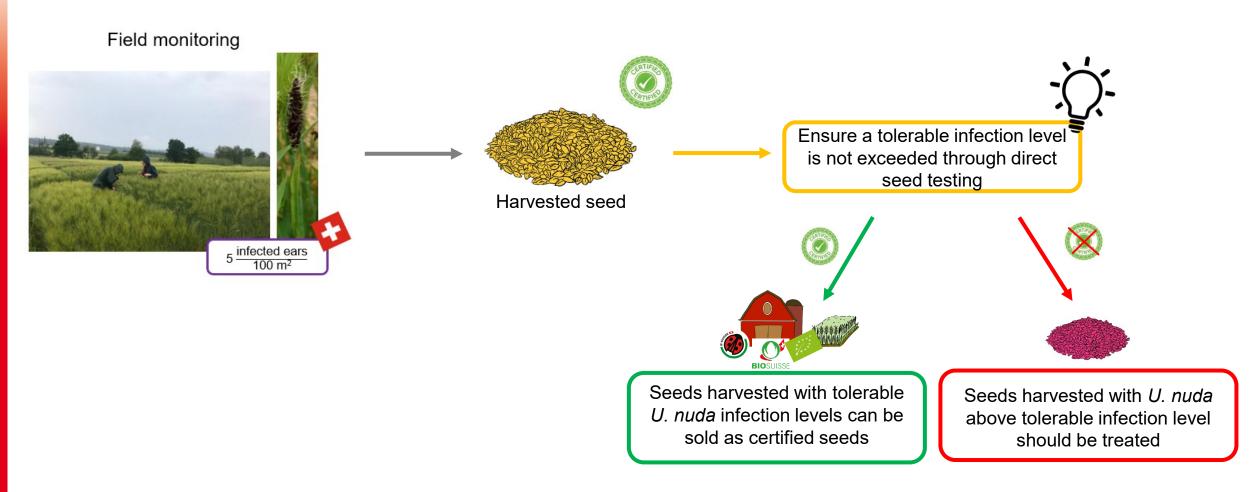


A healthy field can be infected by a neighboring, uncontrolled field



Not applicable on a large scale

Solution for field monitoring problems



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Direct seed inspection

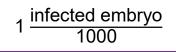
Ensure a tolerable infection level is not exceeded through direct seed testing



There is a method, but it is not often used



Visual laboratory examination of extracted embryos (embryo test)



35 infected ears 100 m²



Problems with this method:



Time-consuming and laborious

The extraction of at least 1000 embryos and their individual testing for infections is required



Infections are difficult to detect

The mycelia in the embryo may be small and barely visible



Not applicable on a large scale

An inspector checks an average of 4 samples per day

© Possible solution for direct seed inspection problems

Aims

- Time savings
- Reduction in errors
- Precise detection of infections
- Development of a scalable method



Confirmation of field monitoring certification with *U. nuda* molecular detection on harvested seeds



With a molecular method!

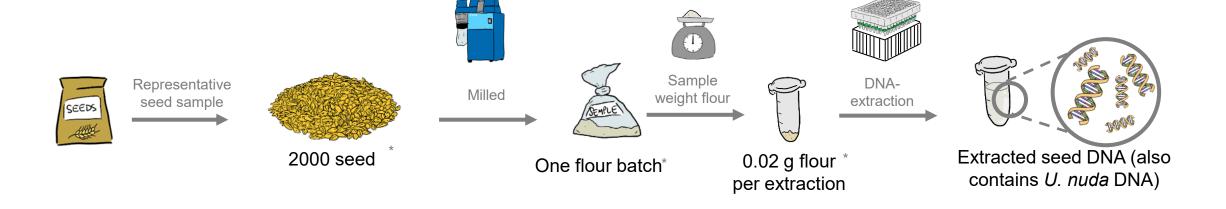
Advantages of molecular methods:

Detects low levels of infection

Tests more than a hundred samples in a week



Our *U. nuda* qPCR method



Our *U. nuda* qPCR method



Analysis of extracted seed DNA + primer and probe using multiplex qPCR method

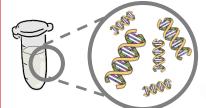
The probes are small glowing "torches" that only switch on when they find the piece of DNA they are looking for



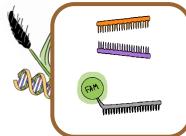
The light emitted by the probes for detecting barley is similar in all samples



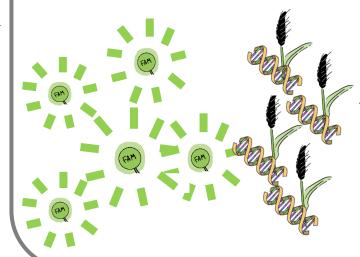
Hordeum vulgare primers



Extracted seed DNA



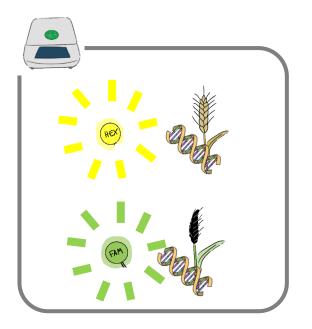
Newly design primers and probes specifically for barley and *U. nuda*



The total amount of light emitted by the probes for *U. nuda* varys depending on the degree of infection

More light captured indicates higher infection

Our *U. nuda* qPCR method



The emitted light is used to quantify U. nuda and barley DNA in each seed sample

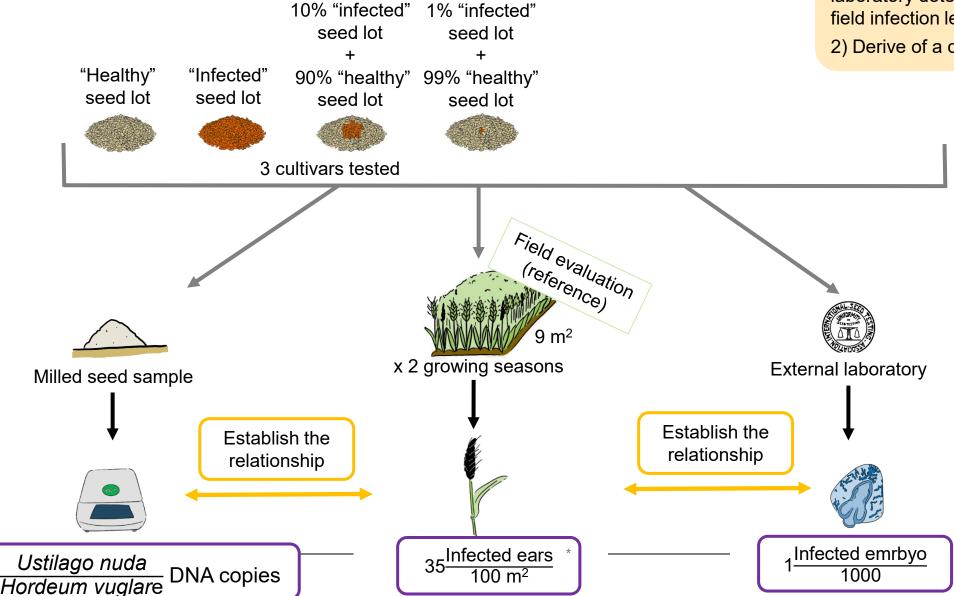


The infection for each sample is quantified by the ratio between the measured *U. nuda* DNA to the measured barley DNA

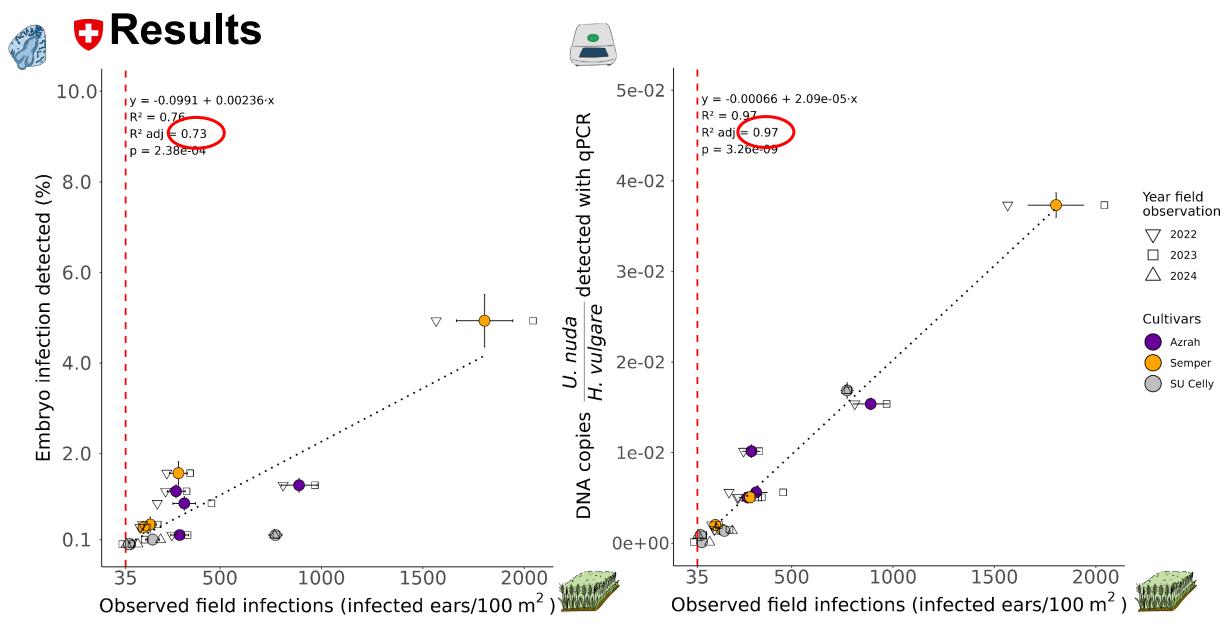


Check correlation between infection detected by qPCR and field infections in plants from the same seed!

Ustilago nuda seed trial



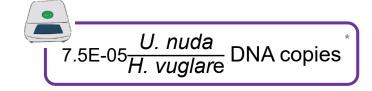
- 1) Establish the relationship between laboratory detection methods and the field infection level
- 2) Derive of a qPCR tolerance threshold

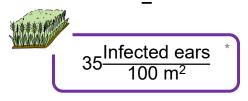


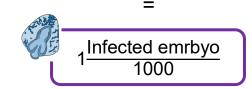
The infection detected by qPCR better reflects the infections observed in the field than the infection determined by the embryo test

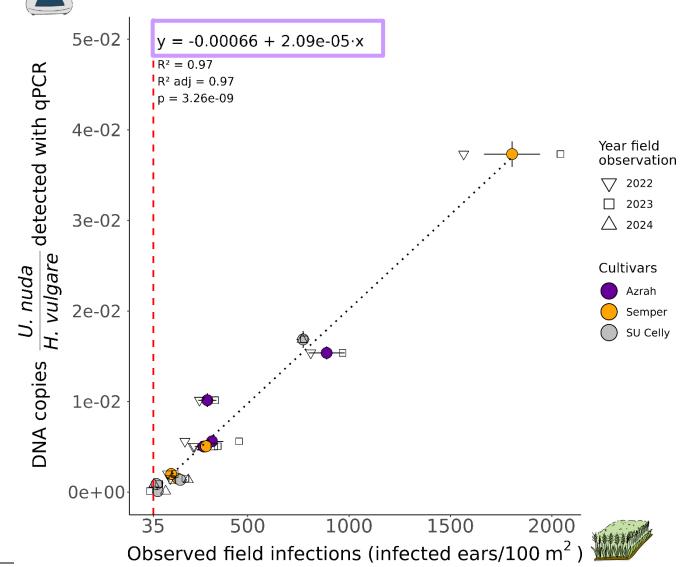
♥Results

Tolerance thresholds



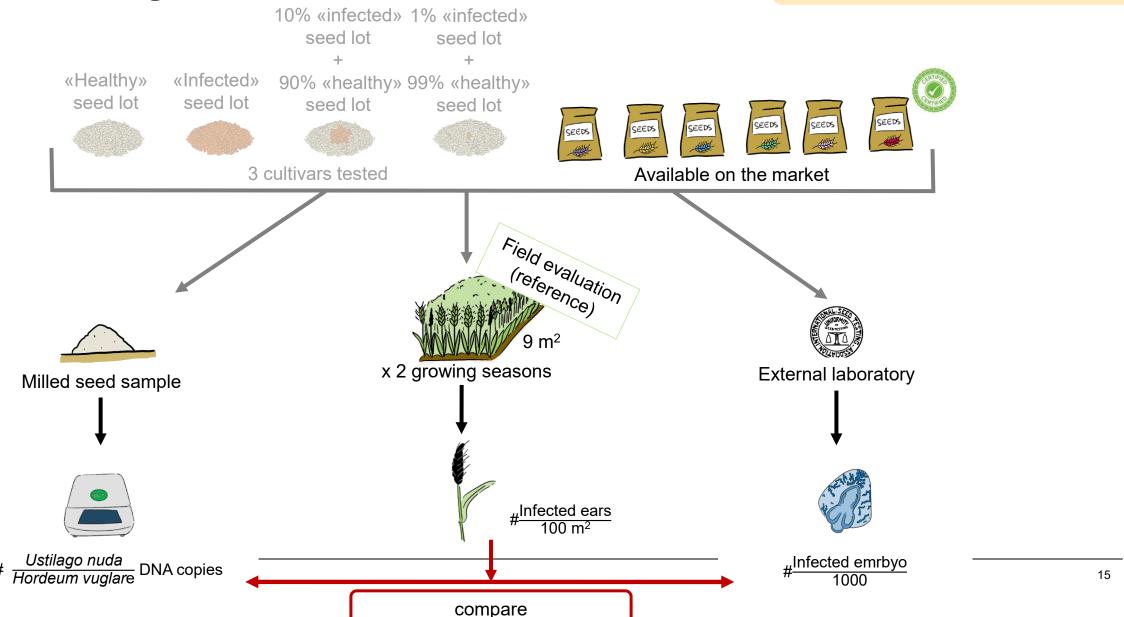






Ustilago nuda commercial seed trial

Aim: determine the best laboratory detection method



Ustilago nuda commercial seed trial: results

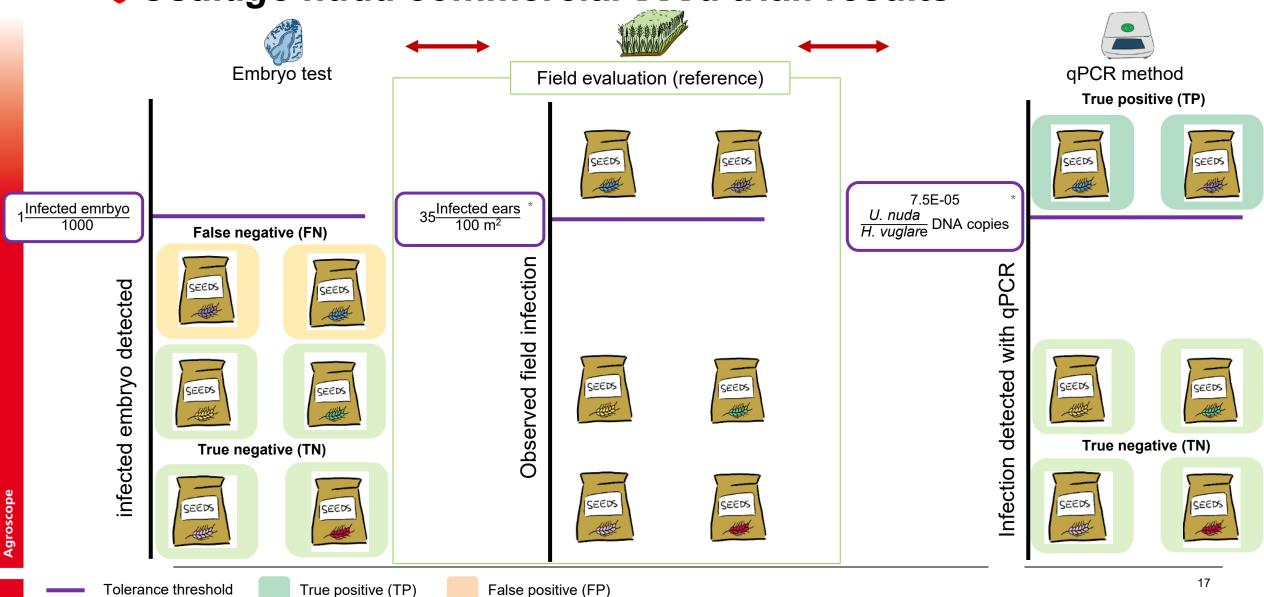
Seed lots with infection above the tolerance threshold Tolerance threshold Infection detected Seed lots with infection below the tolerance threshold

Ustilago nuda commercial seed trial: results

False negative (FN)

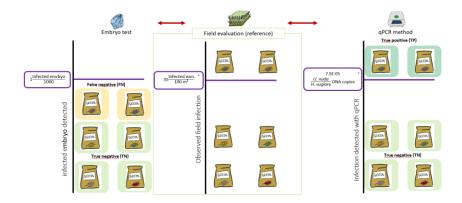
Infection detected

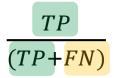
True negative (TN)

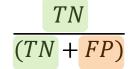


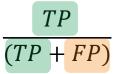
Agroscope

Ustilago nuda commercial seed trial: results









$$\frac{TP + TN}{(TP + TN + FP + FN)}$$

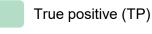
| Laboratory detection methods | Sensitivity | Specificity | Precision | Accuracy |
|------------------------------|-------------|-------------|-----------|-----------|
| Embryo test | SEEDS | SEEDS | SEEDS = | SSEEK (S) |
| qPCR method | | | | SCEEDS = |

Sensitivity measures how well a method recognizes infected seed lots

Specificity measures how well a method recognizes uninfected seed lots Precision measures how likely a positive test result is actually infected

Accuracy measures how well a method correctly classifies both infected and non-infected seed lots

Tolerance threshold Infection detected



True negative (TN)

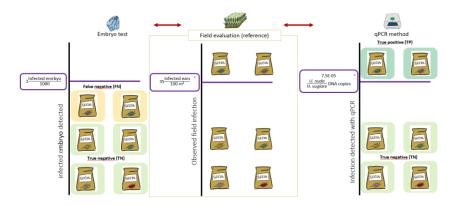


False positive (FP)



False negative (FN)

Ustilago nuda commercial seed trial: results



TP(TP+FN)

TN(TN + FP)

TP(TP + FP)

$$\frac{TP + TN}{(TP + TN + FP + FN)}$$

| Laboratory detection methods | Sensitivity | Specificity | Precision | Accuracy |
|------------------------------|-------------|-------------|-----------|----------|
| Embryo test | 0% | 100% | - | 60% |
| qPCR method | 100% | 100% | 100% | 100% |

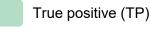
Sensitivity measures how well a method recognizes infected seed lots

Specificity measures how well a method recognizes uninfected seed lots

Precision measures how likely a positive test result is actually infected

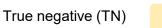
Accuracy measures how well a method correctly classifies both infected and non-infected seed lots

Tolerance threshold Infection detected





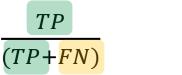
False positive (FP)



False negative (FN)

Agroscope

Ustilago nuda commercial seed trial: results



$$\frac{TN}{(TN + FP)}$$

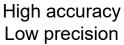
$$\frac{TP}{(TP + FP)}$$

$$\frac{TP + TN}{(TP + TN + FP + FN)}$$

| Laboratory detection methods | Sensitivity | Specificity | Precision | Accuracy |
|------------------------------|-------------|-------------|-----------|----------|
| Embryo test | 0% | 100% | - | 60% |
| qPCR method | 100% | 100% | 100% | 100% |

qPCR method successfully identifies seed lots with Infection either below or above the field tolerance threshold



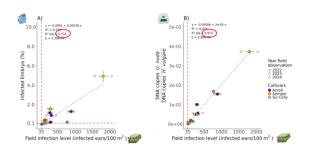




High accuracy High precision

USummary

 The infection detected by qPCR better reflects the infections observed in the field than the infection determined by the embryo test.



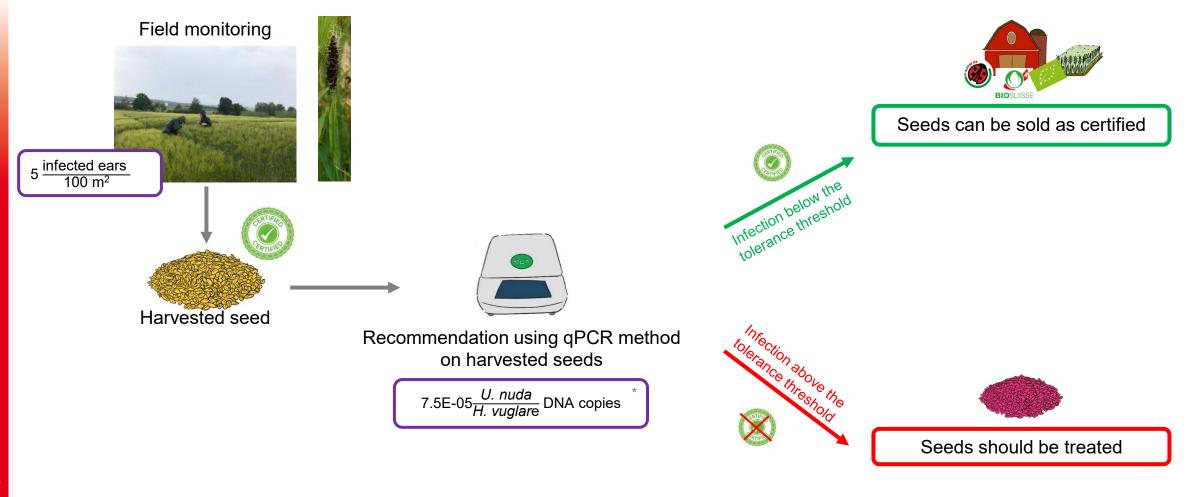
 Our qPCR method accurately classifies samples as either above or below the tolerance threshold matching with the reference

| | $\frac{RP}{(RP + FN)}$ | $\frac{RN}{(RN + FP)}$ | $\frac{RP}{(RP + FP)}$ | $\frac{RP + RN}{(RP + RN + FP + FN)}$ |
|----------------|------------------------|------------------------|------------------------|---------------------------------------|
| Labor-Methoden | Sensitivität | Spezifität | Präzision | Genauigkeit |
| Embryotest 🎆 | 0% | 100% | - | 60% |
| qPCR Methode | 100% | 100% | 100% | 100% |

 Overall, the developed qPCR method is a better predictor of fire blight infections in the field, especially in cases of low infection, compared to the embryo test.



Conclusion



Agroscope

Acknowledgements



Supervisor: Karen Sullam Molecular Ecology



Agroscope research Groups:

Academic Supervisor: Daniel Croll Universität Neuchâtel



Molecular Ecology:

- Franco Widmer
- Marco Wüthrich
- All the group



Seed quality:

- Thomas Hebeisen
- Nicole Bischofberger
- Damian Amrein



Extension arable crops:

- Susanne Vogelgsang
- Irene Bänziger
- Eveline Jenny
- Andreas Kägi
- Francesco Bassi
- Magnus Wagner



External researchers:

The Bavarian State Research Center for Agriculture (LfL):

Peter Büttner

Funding:











