

Editorial

Evolving insights to understanding mycorrhizas

Almost all land plant species form a symbiosis with mycorrhizal fungi. These soil fungi provide nutrients and other services to plants in return for plant carbohydrates. The recent application of microbial metagenomics, metatranscriptomics, and metabolomics to plants and their immediate surroundings confirms the key role of mycorrhizal fungi, rhizosphere bacteria and fungi, and suggests a world of hitherto undiscovered interactions (van der Heijden *et al.*, this issue, pp. 1406–1423). This novel knowledge is leading to a paradigm-shifting view: plants cannot be considered as isolated individuals any more, but as metaorganisms, or holobionts (Hacquard & Schadt, this issue, pp. 1424–1430) encompassing an active microbial community re-programming host physiology (see Pozo *et al.*, this issue, pp. 1431–1436). This bears tremendous implications for plant ecophysiology and evolution, plant breeding, crop management and sustainable ecosystem management.

Mycorrhizal associations are centerpieces in this wide cortege of plant-associated soil biota. To exploit these evolving insights, critical gaps need to be filled in our current understanding of mycorrhizal interactions. This special issue of *New Phytologist* addresses fundamental gaps and contains 30 new contributions on mycorrhizal science, covering topics from genomes to ecosystems. These contributions result from the 33rd New Phytologist Symposium *Networks of power and influence: ecology and evolution of symbioses between plants and mycorrhizal fungi* (Zürich, 14–16 May 2014, <http://www.newphytologist.org/symposiums/view/4>; see also Bender *et al.*, 2014).

A series of key questions addressed in this issue explore some of the most cutting-edge approaches, and include: (1) How is the balance of mutualism maintained between plants and fungi? (2) What is the role of mycorrhizal fungi in the soil ecosystem? (3) What controls fungal community composition, and how is diversity maintained? While many of these questions originated in the earliest days of mycorrhizal science (Koide & Mosse, 2004), the development of new tools and approaches, from genomics to mathematical models to isotopes, is allowing them to be addressed in greater detail, clarity and depth than ever before.

The balance of mutualism and partner selection

From the earliest days of mycorrhizal science, a perennial question has been the balance of mutualism and parasitism (Bronstein *et al.*, 2014), particularly in arbuscular mycorrhizas (AMs)

(Koide & Mosse, 2004). As obligate symbionts, the fungal partners in AM always benefit (Smith & Smith, this issue, pp. 1381–1384), but benefits for the plant partner can vary substantially with environmental conditions (Johnson *et al.*, this issue, pp. 1473–1484), and depending on host–plant identity and development. Maintaining symbiosis in the face of the variable benefits received, depends on the ability of plant and fungal partners to select for or against one another (Werner & Kiers (2015a), this issue, pp. 1437–1442), and to avoid interaction with ‘cheaters’, fungi or plants that acquire resources from the mutualism without providing reciprocal rewards. The best approaches to understanding these interactions and explaining plant–fungal coexistence remain unclear. Werner & Kiers (2015a) argue that these processes can be best understood by the biological market theory (Noe & Hammerstein, 1994), where each individual can change partner on the basis of benefits supplied by other partners, as in human market economy. While not explicitly using market theory, Engelmoer & Kiers (this issue, pp. 1485–1491) provide, using *in vitro* root cultures, an elegant example of how a market prediction, that is, that plants connected in a mycorrhizal network should reduce investment to avoid benefiting competing plants, can be directly tested. Despite being based on relatively simple market analogies, this provides a novel understanding of mycorrhizal hyphal networks from a fungal view. Further work should now test carbon allocation to mycorrhizal networks in plant–soil systems where plant carbohydrate availability differs from those in *in vitro* root cultures.

While the market analogy can clearly serve a purpose, Smith & Smith and van der Heijden *et al.* raise important cautions about extending the market analogy for mycorrhizal mutualisms beyond the discussion of general principles. There are a number of important aspects of the mutualism that the market analogy fails to capture. One, raised by Smith & Smith, is the issue of mycoheterotrophs (see Field *et al.*, this issue, pp. 1492–1502), and so-called ‘cheaters’. Indeed, there is evidence for manipulations of host gene expression by colonizing fungal symbionts through effector proteins (Plett *et al.*, 2014), opening the way to manipulation of the market. Bever (this issue, pp. 1503–1514) shows how a mechanistic model helps to provide greater insight into these interactions. The simple assumption that plants must be first colonized by fungi before being able to assess their symbiotic efficiency, provides a mechanism whereby less efficient fungal symbionts can persist over a relatively wide range of environmental conditions. Field *et al.* further extend the importance of considering plant–fungal exchanges over the whole lifespan of the association, showing that the mycoheterotrophic fern *Ophioglossum vulgatum* has developed a ‘take now, pay later’ strategy, being entirely dependent on fungi for both nutrients and carbon as a gametophyte and young sporophyte, but potentially repaying this investment as mature sporophytes.

There are a number of additional concerns about the market analogy and the application of economic theory to symbioses. First, the application of market theory to mycorrhizal symbioses has frequently assumed a two-goods market, involving carbon and phosphorus. While some have argued that different types of resources (nitrogen, phosphorus, carbon) are analogous to multiple different currencies in markets (Bloom *et al.*, 1985), this analogy fails to recognize two aspects of plant physiology. First, mycorrhizal benefits are diverse for the plant (Selosse & Rousset, 2011), and plant protection by mycorrhizal fungi is also relevant (Newsham *et al.*, 1995): for example, in this issue, Pozo *et al.* examine the mechanisms by which mycorrhizas shape plant defense against pathogens and stress. Second, one essential nutrient simply cannot serve the same purpose or be exchanged for another: this fundamental principle underpins 'Liebig's law of the minimum' (Johnson, 2010), which has, despite some limitations, become one of the foundations of environmental stoichiometry. In this issue Johnson *et al.* integrate Liebig's law and stoichiometric theory with plant–fungal interactions in mycorrhizas, showing how the carbon:phosphorus exchange between plants and AM fungi breaks down when nitrogen is the main limiting nutrient.

Another major limitation of market theory is that the fungal community associating with a plant does not simply respond to plant resource allocation under market rules, but is also shaped by direct antagonistic competition among fungal species, as well as interactions with fungivores, pathogens of fungi, other soil biota and stochastic events. One example of this is the effect of assembly history, where the order of fungal species' arrival into an ecosystem determines community composition (Kennedy & Bruns, 2005; Dickie *et al.*, 2012; Peay *et al.*, 2012). This has also been shown for AM, where the fungal species that colonizes the root system first is able to effectively prevent or reduce colonization by later fungi (Abbott & Robson, 1984; Werner & Kiers (2015b), this issue, pp. 1515–1524). Werner & Kiers (2015b; see also Commentary by Johnson *et al.*, in this issue, pp. 1375–1377) discuss preferential allocation by hosts as potential mechanisms, but it is notable that other fungal systems also show priority effects in the absence of any host allocation (e.g. Dickie *et al.*, 2012; Peay *et al.*, 2012). This may imply that we should give more consideration to competition and direct fungal–fungal antagonism in mycorrhizal fungi; a mechanism that falls well outside traditional market theory or the mechanistic models of Bever. Finally, most current works use growth or nutrient flow as markers for the mutualism, but these are only proxies: in the future, a more evolutionary-relevant view of mutualism (or cheating) should ideally investigate the partners' fitness, that is, the ability to survive and reproduce.

Role in soil carbon

The carbon allocated by plants to mycorrhizas does not just support the fungus, but also has potentially profound effects on the soil ecosystem. Up to 50% of plant carbon is allocated to mycorrhizal fungi, pointing to the key role of mycorrhizal fungi in the carbon cycle (van der Heijden *et al.*). The recent finding that carbon sequestration in soil is related to mycorrhizal fungal community shifts during succession further points to the key role of mycorrhizal

fungi for the carbon cycle (Clemmensen *et al.*, this issue, pp. 1525–1536; see also the Commentary by Fernandez & Kennedy, this issue, pp. 1378–1380). A particularly vexing question in mycorrhizal ecology is the question of how carbon allocation to mycorrhizal fungi influences decomposition and the interaction of mycorrhizal fungi with free-living saprotrophs and litter decomposition. Using NanoSIMS imaging, Kaiser *et al.* (this issue, pp. 1537–1551) show the remarkably fast transfer of carbon from host plants to mycorrhizal hyphae, and out into the soil. The exudation of carbon by hyphae, rather than passive root exudation, may allow for much greater precision in using plant photosynthate to facilitate saprotrophic release of organic matter (the so-called priming effect).

Carbon exudation is not the only major pathway of mycorrhizal influence on soil carbon, as mycorrhizal fungi can both participate directly in decomposition (Lindahl & Tunlid, this issue, pp. 1443–1447), and form a substantial carbon pool in their own necromass (Clemmensen *et al.*; Fernandez & Kennedy). These effects are context dependent. In young, relatively nutrient-rich soils the net effect of mycorrhizal fungi on decomposition may be positive through priming and provision of an easily decomposable biomass. In older soils, on the contrary, the net effect of mycorrhizas may be to slow decomposition through the removal of nutrients, which become limiting for other microbes, and more decay-resistant hyphae (Orwin *et al.* 2011; Clemmensen *et al.*). Indeed, an important emerging theme in mycorrhizal research is the importance of fungal traits in predicting effects on ecosystem function (Clemmensen *et al.*; Rillig *et al.*, this issue, pp. 1385–1388).

While significant amounts of carbon pass through mycorrhizal networks, the direct contribution of mycorrhizas to decomposition and the breakdown of organic matter is still poorly documented. The repeated, independent evolutions to the mycorrhizal lifestyle in all fungal lineages are associated with a massive loss of lignocellulose-degrading genes compared to saprotrophic ancestors (Plett & Martin, 2011; van der Heijden *et al.*), suggesting a limiting role for decomposition. However, it appears that several ectomycorrhizal fungi potentially decompose organic matter through the oxidative cleavage of glycosidic bonds in cellulose and hemicellulose, and the oxidative modification and arrangement of lignin upon attack by highly destructive oxygen reactive species (Rineau *et al.*, 2013). They do not acquire carbon from this decomposing activity but mobilize nitrogen compounds embedded in organic matter thanks to a constant host carbohydrate supply (Lindahl & Tunlid). Acquisition of bacterial genes coding for decomposing enzymes through horizontal gene transfer can complement the enzyme arsenal of symbionts, and likely played a role in the evolution of mycorrhizal symbioses (Chaib De Mares *et al.*, this issue, pp. 1552–1564).

Communities and ecosystems

Another long-standing question in mycorrhizal science has been what determines the diversity, structure and composition of fungal communities. Molecular methods, allowing easier and faster detection and identification of fungi, are now driving a revolution in our understanding of fungal communities. Lindahl *et al.* (2013) compiled a user's guide for fungal community ecologists who use

amplicon-based next-generation sequencing (NGS), outlining many of the aspects of NGS technologies that need to be addressed to prevent making biased conclusions. In this issue, Nguyen *et al.* (pp. 1389–1393), follow on the theme, targeting the various aspects of contamination and need for controls in sample handling and NGS data production practices. The awareness of the problems outlined by them, with solutions offered, are of increasing importance as the data volume increases in line with the evolving NGS platforms.

The drivers of community dynamics of mycorrhizal fungi are only partly understood. Mycorrhizal fungal species distribution is influenced by external abiotic and biotic forces, and intrinsic fungal properties (Chaudhary *et al.*, 2008), but the role of biotic interactions (with host, other fungi and any other organisms) in shaping mycorrhizal fungal communities remains poorly investigated, and particularly little is known about co-variation of host and mycorrhizal fungal communities. Zobel & Öpik (2014) have proposed that in primary succession plant communities drive AM fungal communities (Passenger hypothesis), and in secondary succession (including in response to land use) the AM fungal community drives the plant community (Driver hypothesis). In successional stable ecosystems both AM fungal and plant communities co-respond to local environmental gradients (Habitat hypothesis), but for large scales the two communities may have independent dynamics (Independence hypothesis). Empirical evidence is surprisingly scarce on these scenarios. Martínez-García *et al.* (this issue, pp. 1565–1576) now provide evidence that AM fungal communities across long-term ecosystem age gradients, from succession to retrogression, are strongly determined by ecosystem age and structured by host identity, thus providing support to the Passenger and/or Driver hypotheses.

AM fungal community responses to different land uses have been a topic of considerable interest since the demonstration of a dramatic decrease in AM fungal diversity in intensively managed arable fields in comparison to surrounding native vegetation (Helgason *et al.*, 1998). Vályi *et al.* (this issue, pp. 1577–1586) demonstrate that both land use and host plant identity affect AM fungal richness and community composition in a grassland system. Interestingly, the communities in this study were somewhat more taxon rich under high land-use intensity, which is counterintuitive considering the expected negative effect of land use. In agreement with this result, other managed ecosystems have turned out to be not necessarily depauperate in fungal species, both at local (Hijri *et al.*, 2006; Moora *et al.*, 2014) and larger regional scales (Xiang *et al.*, 2014). These results suggest that the response of AM fungal communities to the nature and intensity of land use may vary in direction, possibly in a context-dependent manner. The information we have on AM fungal diversity–disturbance relationships may represent different portions of the land-use intensity gradient. It may turn out that moderate disturbance supports the highest community diversity of these fungi while extremely low and extremely high disturbance negatively affects diversity, as has been proposed for other organisms by the ‘intermediate disturbance hypothesis’ (Hughes *et al.*, 2007).

The influence of abiotic drivers of mycorrhizal fungal communities remains idiosyncratic across studies. Mundra *et al.* (this issue,

pp. 1587–1597) found that while vegetation was structured by abiotic environmental parameters, the fungal communities associated with *Bistorta vivipara* roots were not. Bahram *et al.* (this issue, pp. 1454–1463) present a meta-analysis of mycorrhizal fungal communities of different types across temporal and different spatial scales, including vertical scales of soil depth. Perhaps surprisingly, they could not reveal clear trends except between top- and deeper-soil horizons. This further illustrates our fragmentary knowledge of mycorrhizal fungal diversity patterns at large scales, and suggests context-dependent, rather than uniform processes. Our ability, in the future, to gain clear views of fungal functional traits (Rillig *et al.*) will help the investigation of the processes acting in the establishment of fungal communities.

There are also important temporal components to mycorrhizal fungal communities, as illustrated by Cotton *et al.*'s (this issue, pp. 1598–1607) report of large interannual shifts in AM fungal communities in a soybean cropping system, which contrasts with the more limited interannual turnover observed in less disturbed ecosystems (Bennett *et al.*, 2013). Smaller, seasonal, shifts in orchid mycorrhizal fungal communities are reported by Oja *et al.* (this issue, pp. 1608–1618), whereas habitat and host species were stronger drivers of diversity patterns of these fungi in Estonian meadow and forest ecosystems. This study elegantly complements other recent insights into orchid mycorrhizal fungal community dynamics revealing host-related, spatial and temporal diversity patterns (Jacquemyn *et al.*, 2014; Ercole *et al.*, 2015) which were published in this journal after the 31st New Phytologist Symposium *Orchid symbioses: models for evolutionary ecology* (14–16 May 2013; Selsos, 2014; Bronstein *et al.*, 2014; <http://www.newphytologist.org/symposiums/view/3>).

Among the intrinsic properties of fungi, dispersal properties are deservedly gaining increasing interest. Dispersal limitation – the fact that not all suitable habitats are colonized by a species – has received increasing attention (e.g. Peay & Bruns, 2014). Another aspect of propagule availability is the pool of locally available, but not currently active organisms. In analogy with the plant seed bank concept, fungal spore banks (or propagule banks) in soil represent fungi that are currently dormant, but have the potential to start growing when suitable conditions appear. In this issue, Glassman *et al.* (pp. 1619–1631), provide evidence that ectomycorrhizal fungal spore banks represent a small subset of the mature forest's fungal community and are location dependent.

Conclusion: enlarging the mycorrhizal symbiosis to new players

Mycorrhizal symbiosis turns out to be a more complex network of interaction than often considered because new players enter the field. Lekberg *et al.* (this issue, pp. 1399–1403) challenge the status of nonmycorrhizal plants, suggesting that they could be relevant for AM fungi that colonize them. On the fungal side a new taxon was recently discovered to potentially join the scene: members of the Mucoromycotina were found to form associations with basal nonvascular land plants such as liverworts (Field *et al.*), sometimes together with AM fungi. Rimington *et al.* (this issue, pp. 1394–1398) now provide evidence that a wide range of basal vascular land

plants (ferns and lycopods) also associate with both AM and Mucoromycotina fungi. This is congruent with the recently described dual association with AM and Mucoromycotina fungi in a Devonian fossil plant (Strullu-Derrien *et al.*, 2014) whose phylogenetic position falls between liverworts and vascular plants. The colonization of Mucoromycotina, although obviously biotrophic (symptomless), raises the question of the physiological outcome on the plant side: although beneficial effects of Mucoromycotina have recently been demonstrated for liverworts (Field *et al.*), it deserves studies on more plant models, especially to test whether simultaneous colonization by both groups of fungi provide additional benefits. Bacterial players also enter the scene here: *Endogone*, a Mucoromycotina fungus, contains endosymbiotic bacteria (Desirò *et al.*, this issue, pp. 1464–1472), exactly as found in AM fungi (Ghignone *et al.*, 2012), pointing to further similarities between these two lineages. The role, if any, of these bacteria in mycorrhizas also deserves further study.

On the plant side, the mycorrhizal symbiosis is increasingly viewed as an ecological network, where shared fungal partners create a common mycorrhizal hyphal network (Bender *et al.*, 2014) making each plant an indirect partner of its neighbors. Nutrient transfers are well known, as well as the imbalanced contribution of plant partners to mycorrhizal networks as compared to the benefit they gain, although the determinism for such outcome remains unclear (Walder *et al.*, this issue, pp. 1632–1645). New functions are now discovered: plant defense signals can be transferred from one plant to another through mycorrhizal networks as conduits (Johnson & Gilbert, this issue, pp. 1488–1493). Plants connected to networks are subsequently better protected against insect herbivores and antagonistic herbivores. Although the mechanisms, and the evolutionary forces that shape such indirect collaborations remain unclear, it is now evident that the mycorrhizal fungal community blurs the limits of plant holobionts.

Taken as a whole, the articles in this special issue represent a remarkable suite of studies, all focused on mycorrhizal science but ranging in scale from the molecular to covering whole-ecosystems. While the questions they address have a long legacy, the application of novel techniques and ideas has allowed significant advances in the understanding of this complex symbiosis. Mycorrhizal science has, perhaps more than many other fields of research, remained an integrated field; pulling together approaches from systematics, molecular biology, soil science, and ecology. This represents a significant strength of the field, and one that *New Phytologist* as a nonprofit trust has striven to support for many decades, in the pages of the journal and through symposia and workshops, in a lasting and deep-rooted 'love affair' (Selosse & Martin, 2013). *New Phytologist* will attend the 8th International Conference of Mycorrhiza (ICOM8), 3–7 August 2015, Flagstaff, AZ, USA, which represents the next step in this long-standing association.

Acknowledgements

I.A.D. is supported by the Bio-Protection Research Centre, Lincoln University, New Zealand. F.M.M. is supported by the Laboratory of Excellence ARBRE (ANR-11-LABX-0002-01) and

the Genomic Science Program (Plant–Microbe Interactions project) funded by the US Department of Energy (Contract DE-AC05-00OR22725). M.Ö. is funded by the Estonian Research Council (grants 9050, IUT20-28) and the European Regional Development fund (Centre of Excellence FIBIR).

Ian A. Dickie¹, Ian Alexander², Sarah Lennon³, Maarja Öpik⁴, Marc-André Selosse⁵, Marcel G. A. van der Heijden⁶ and Francis M. Martin^{7*}

¹Bio-Protection Research Centre, Lincoln University, PO Box 85084, Lincoln 7647, New Zealand;

²School of Biological Sciences, University of Aberdeen, Cruickshank Building, Aberdeen, AB24 3UU, UK;

³New Phytologist Central Office, Lancaster University, Bailrigg House, Lancaster, LA1 4YE, UK;

⁴Department of Botany, Institute of Ecology and Earth Sciences, University of Tartu, 40 Lai St, 51005 Tartu, Estonia;

⁵Département Systématique et Evolution, Muséum national d'Histoire naturelle, UMR 7205 ISYEB CP 50 45 rue Buffon, Paris 75005, France;

⁶Agroscope – Institute for Sustainability Sciences, Reckenholzstrasse 191, CH-8046 Zurich, Switzerland;

⁷Lab of Excellence ARBRE, INRA, UMR 1136 INRA-Université de Lorraine 'Interactions Arbres/Micro-organismes', 54280 Champenoux, France

(*Author for correspondence: tel +33 383 39 40 80; email: fmartin@nancy.inra.fr)

References

- Abbott LK, Robson AD. 1984. Colonization of the root system of subterranean clover by three species of vesicular-arbuscular mycorrhizal fungi. *New Phytologist* 96: 275–281.
- Bahram M, Peay KG, Tedersoo L. 2015. Local-scale biogeography and spatiotemporal variability in communities of mycorrhizal fungi. *New Phytologist* 205: 1454–1463.
- Bender SF, da Silva Valadares RB, Taudiere A. 2014. Mycorrhizas: dynamic and complex networks of power and influence. *New Phytologist* 204: 15–18.
- Bennett AE, Daniell TJ, Öpik M, Davison J, Moora M, Zobel M, Selosse MA, Evans D. 2013. Arbuscular mycorrhizal fungal networks vary throughout the growing season and between successional stages. *PLoS ONE* 8: e83241.
- Bever JD. 2015. Preferential allocation, physio-evolutionary feedbacks, and the stability and environmental patterns of mutualism between plants and their root symbionts. *New Phytologist* 205: 1503–1514.
- Bloom AJ, Chapin FS, Mooney HA. 1985. Resource limitation in plants – an economic analogy. *Annual review of Ecology and Systematics* 16: 363–392.
- Bronstein JL, Armbruster WS, Thompson JN. 2014. Understanding evolution and complexity of species interactions using orchids as a model system. *New Phytologist* 202: 373–375.
- Chaib De Mares M, Hess J, Floudas D, Lipzen A, Choi C, Kennedy M, Grigoriev IV, Pringle A. Horizontal transfer of carbohydrate metabolism genes into ectomycorrhizal *Amanita*. *New Phytologist* 205: 1552–1564.
- Chaudhary VB, Lau MK, Johnson NC. 2008. Macroecology of microbes – biogeography of the Glomeromycota. In: Varma A, ed. *Mycorrhiza: genetics and molecular biology, eco-function, biotechnology, eco-physiology, structure and systematics*. Berlin/Heidelberg, Germany: Springer-Verlag, 529–562.
- Clemmensen KE, Finlay RD, Dahlberg A, Stenlid J, Wardle DA, Lindahl BD. 2015. Carbon sequestration is related to mycorrhizal fungal community shifts during long term succession in boreal forests. *New Phytologist* 205: 1525–1536.

- Cotton TEA, Fitter AH, Miller RM, Dumbrell AJ, Helgason T. 2015. Fungi in the future: interannual variation and effects of atmospheric change on arbuscular mycorrhizal fungal communities. *New Phytologist* 205: 1598–1607.
- Desirò A, Faccio A, Kaech A, Bidartondo MI, Bonfante P. 2015. *Endogone*, one of the oldest plant-associated fungi, host unique Mollicutes-related endobacteria. *New Phytologist* 205: 1464–1472.
- Dickie IA, Fukami T, Wilkie JP, Allen RB, Buchanan PK. 2012. Do assembly history effects attenuate from species to ecosystem properties? A field test with wood-inhabiting fungi. *Ecology Letters* 15: 133–141.
- Engelmoer DJP, Kiers ET. 2015. Host diversity affects the abundance of the extraradical arbuscular mycorrhizal network. *New Phytologist* 205: 1485–1491.
- Ercole E, Adamo M, Rodda M, Gebauer G, Girlanda M, Perotto S. 2015. Temporal variation in mycorrhizal diversity and carbon and nitrogen stable isotope abundance in the wintergreen meadow orchid *Anacamptis morio*. *New Phytologist* 205: 1308–1319.
- Fernandez CW, Kennedy PG. 2015. Moving beyond the black-box: fungal traits, community structure, and carbon sequestration in forest soils. *New Phytologist* 205: 1378–1380.
- Field KJ, Leake Jr, Tille S, Allinson KE, Rimington WR, Bidartondo MI, Beerling DJ, Cameron DD. 2015. From mycoheterotrophy to mutualism: mycorrhizal specificity and functioning in *Ophioglossum vulgatum* sporophytes. *New Phytologist* 205: 1492–1502.
- Ghignone S, Salvioli A, Anca I, Lumini E, Ortu G, Petiti L, Cruveiller S, Bianciotto V, Piffanelli P, Lanfranco L *et al.* 2012. The genome of the obligate endobacterium of an AM fungus reveals an interphylum network of nutritional interactions. *ISME Journal* 6: 136–45.
- Glassman SI, Peay KG, Talbot JM, Smith DP, Chung JA, Taylor JW, Vilgalys R, Bruns TA. 2015. continental view of pine-associated ectomycorrhizal spore banks: a quiescent functional guild with a strong biogeographic pattern. *New Phytologist* 205: 1619–1631.
- Hacquard S, Schadt CW. 2015. Towards a holistic understanding of the beneficial interactions across the *Populus* microbiome. *New Phytologist* 205: 1424–1430.
- van der Heijden MGA, Martin F, Selosse M-A, Sanders IR. 2015. Mycorrhizal ecology and evolution: the past, the present, and the future. *New Phytologist* 205: 1406–1423.
- Helgason T, Daniell TJ, Husband R, Fitter AH, Young JPW. 1998. Ploughing up the wood-wide web? *Nature* 394: 431.
- Hijiri I, Sykora Z, Oehl F, Ineichen K, Mäder P, Wiemken A, Redecker D. 2006. Communities of arbuscular mycorrhizal fungi in arable soils are not necessarily low in diversity. *Molecular Ecology* 15: 2277–2289.
- Hughes AR, Byrnes JE, Kimbro DL, Stachowicz JJ. 2007. Reciprocal relationships and potential feedbacks between biodiversity and disturbance. *Ecology Letters* 10: 849–864.
- Jacquemyn H, Brys R, Merckx VSFT, Waud M, Lievens B, Wiegand T. 2014. Coexisting orchid species have distinct mycorrhizal communities and display strong spatial segregation. *New Phytologist* 202: 616–627.
- Johnson D. 2015. Priorities for research on priority effects. *New Phytologist* 205: 1375–1377.
- Johnson D, Gilbert L. 2015. Interplant signalling through hyphal networks. *New Phytologist* 205: 1448–1453.
- Johnson NC, Wilson GWT, Wilson JA, Miller RM, Bowker MA. 2015. Mycorrhizal phenotypes and the Law of the Minimum. *New Phytologist* 205: 1473–1484.
- Johnson NC. 2010. Resource stoichiometry elucidates the structure and function of arbuscular mycorrhizas across scales. *New Phytologist* 185: 631–647.
- Kaiser C, Kilburn MR, Clode PL, Fuchslueger L, Koranda M, Cliff JB, Solaiman ZM, Murphy D. 2015. Exploring the transfer of recent plant photosynthates to soil microbes: mycorrhizal pathway vs direct root exudation. *New Phytologist* 205: 1537–1551.
- Kennedy PG, Bruns TD. 2005. Priority effects determine the outcome of ectomycorrhizal competition between two *Rhizopogon* species colonizing *Pinus muricata* seedlings. *New Phytologist* 166: 631–638.
- Koide RT, Mosse B. 2004. A history of research on arbuscular mycorrhiza. *Mycorrhiza* 14: 145–163.
- Lekberg Y, Rosendahl S, Olsson PA. 2015. The fungal perspective of arbuscular mycorrhizal colonization in ‘nonmycorrhizal’ plants. *New Phytologist* 205: 1399–1403.
- Lindahl BD, Tunlid A. 2015. Ectomycorrhizal fungi – potential organic matter decomposers, yet not saprotrophs. *New Phytologist* 205: 1443–1447.
- Lindahl BD, Nilsson RH, Tedersoo L, Abarenkov K, Carlsen T, Kjoller R, Kauserud H. 2013. Fungal community analysis by high-throughput sequencing of amplified markers – a user’s guide. *New Phytologist* 199: 288–299.
- Martínez-García LB, Richardson SJ, Tylianakis JM, Peltzer DA, Dickie IA. 2015. Host identity is a dominant driver of mycorrhizal fungal community composition during ecosystem development. *New Phytologist* 205: 1565–1576.
- Moora M, Davison J, Öpik M, Metsis M, Saks Ü, Jairus T, Vasar M, Zobel M. 2014. Anthropogenic land use shapes the composition and phylogenetic structure of soil arbuscular mycorrhizal fungal communities. *FEMS Microbiology Ecology* 90: 609–621.
- Mundra S, Halvorsen R, Kauserud H, Müller E, Vik U, Eidesen PB. 2015. Arctic fungal communities associated with roots of *Bistorta vivipara* do not respond to the same fine-scale edaphic gradients as the aboveground vegetation. *New Phytologist* 205: 1587–1597.
- Newsham KK, Fitter AH, Watkinson AR. 1995. Multi-functionality and biodiversity in arbuscular mycorrhizas. *Trends in Ecology & Evolution* 10: 407–411.
- Nguyen NH, Smith D, Peay K, Kennedy P. 2015. Parsing ecological signal from noise in next generation amplicon sequencing. *New Phytologist* 205: 1389–1393.
- Noe R, Hammerstein P. 1994. Biological markets: supply and demand determine the effect of partner choice in cooperation, mutualism and mating. *Behavioral Ecology and Sociobiology* 35: 1–11.
- Oja J, Kohout P, Tedersoo L, Kull T, Kõljalg U. 2015. Temporal patterns of orchid mycorrhizal fungi in meadows and forests as revealed by 454 pyrosequencing. *New Phytologist* 205: 1608–1618.
- Orwin KH, Kirschbaum MU, St John MG, Dickie IA. 2011. Organic nutrient uptake by mycorrhizal fungi enhances ecosystem carbon storage: a model-based assessment. *Ecology Letters*, 14: 493–502.
- Peay KG, Belisle M, Fukami T. 2012. Correction for phylogenetic relatedness predicts priority effects in nectar yeast communities. *Proceedings. Biological sciences/The Royal Society* 279: 5066.
- Peay KG, Bruns TD. 2014. Spore dispersal of basidiomycete fungi at the landscape scale is driven by stochastic and deterministic processes and generates variability in plant–fungal interactions. *New Phytologist* 204: 180–191.
- Plett JM, Martin F. 2011. Blurred boundaries: lifestyle lessons from ectomycorrhizal fungal genomes. *Trends in Genetics* 27: 14–22.
- Plett JM, Daguerre Y, Wittulski S, Vayssières A, Deveau A, Melton SJ, Kohler A, Morrell-Falvey J, Brun A, Veneault-Fourrey C *et al.* 2014. The effector MiSSP7 of the mutualistic fungus *Laccaria bicolor* stabilizes the *Populus* JAZ6 protein and represses JA-responsive genes. *Proceedings of the National Academy of Sciences, USA* 111: 8299–8304.
- Pozo MJ, Lopez-Raez JA, Azcón-Aguilar C, García-Garrido JM. 2015. Phytohormones as integrators of environmental signals in the regulation of mycorrhizal symbioses. *New Phytologist* 205: 1431–1436.
- Rillig MC, Aguilar-Trigueros CA, Bergmann J, Verbruggen E, Veresoglou SD, Lehmann A. 2014. Plant root and mycorrhizal fungal traits for understanding soil aggregation. *New Phytologist* 205: 1385–1388.
- Rimington WR, Pressel S, Duckett JG, Bidartondo MI. 2015. Fungal associations of basal vascular plants: reopening a closed book. *New Phytologist* 205: 1394–1398.
- Rineau F, Shah F, Smits MM, Persson P, Johansson T, Carleer R, Troein C, Tunlid A. 2013. Carbon availability triggers the decomposition of plant litter and assimilation of nitrogen by an ectomycorrhizal fungus. *ISME Journal* 10: 2010–22.
- Selosse M-A. 2014. The latest news from biological interactions in orchids: in love, head to toe. *New Phytologist* 202: 337–340.
- Selosse M-A, Martin F. 2013. Mycorrhizas and *New Phytologist*: une vraie histoire d’amour. *New Phytologist* 200: 587–589.
- Selosse M-A, Rousset F. 2011. The plant–fungal marketplace. *Science* 333: 828–829.
- Smith FA, Smith SE. 2015. How harmonious are arbuscular mycorrhizal symbioses? Inconsistent concepts reflect different mindsets as well as results. *New Phytologist* 205: 1381–1384.
- Strullu-Derrien C, Kenrick P, Pressel S, Duckett JG, Rioult JP, Strullu DG. 2014. Fungal associations in *Horneophyton ligneri* from the Rhynie Chert (c. 407 million

- year old) closely resemble those in extant lower land plants: novel insights into ancestral plant–fungus symbioses. *New Phytologist* **203**: 964–979.
- Vályi K, Rillig MC, Hempel S. 2015. Land-use intensity and host plant identity interactively shape communities of arbuscular mycorrhizal fungi in roots of grassland plants. *New Phytologist* **205**: 1577–1586.
- Walder F, Brulé D, Koegel S, Wiemken A, Boller T, Courty P-E. 2015. Plant phosphorus acquisition in a common mycorrhizal network: regulation of phosphate transporter genes of the Pht1 family in sorghum and flax. *New Phytologist* **205**: 1632–1645.
- Werner G, Kiers ET. 2015a. Partner selection in the mycorrhizal symbiosis. *New Phytologist* **205**: 1437–1442.
- Werner G, Kiers ET. 2015b. Order of arrival structures arbuscular mycorrhizal colonization of plants. *New Phytologist* **205**: 1515–1524.
- Xiang D, Verbruggen E, Hu Y, Veresoglou SD, Rillig MC, Zhou W, Chen B. 2014. Land use influences arbuscular mycorrhizal fungal communities in the farming–pastoral ecotone of northern China. *New Phytologist* **204**: 968–978.
- Zobel M, Öpik M. 2014. Plant and arbuscular mycorrhizal fungal (AMF) communities—which drives which? *Journal of Vegetation Science* **25**: 1133–1140.
- Key words:** 33rd New Phytologist Symposium, ecology, evolution, mycorrhizas, symbioses.



About New Phytologist

- *New Phytologist* is an electronic (online-only) journal owned by the New Phytologist Trust, a **not-for-profit organization** dedicated to the promotion of plant science, facilitating projects from symposia to free access for our Tansley reviews.
- Regular papers, Letters, Research reviews, Rapid reports and both Modelling/Theory and Methods papers are encouraged. We are committed to rapid processing, from online submission through to publication 'as ready' via *Early View* – our average time to decision is <26 days. There are **no page or colour charges** and a PDF version will be provided for each article.
- The journal is available online at Wiley Online Library. Visit **www.newphytologist.com** to search the articles and register for table of contents email alerts.
- If you have any questions, do get in touch with Central Office (np-centraloffice@lancaster.ac.uk) or, if it is more convenient, our USA Office (np-usaoffice@lancaster.ac.uk)
- For submission instructions, subscription and all the latest information visit **www.newphytologist.com**