



Priority Effects in the Apple Flower Determine If the Siderophore Desferrioxamine Is a Virulence Factor for *Erwinia amylovora* CFBP1430

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ABSTRACT Iron is crucial for bacterial growth and virulence. Under iron-deficiency bacteria produce siderophores, iron chelators that facilitate the iron uptake into the cell via specific receptors. *Erwinia amylovora*, the causative agent of fire blight, produces hydroxamate-type desferrioxamine siderophores (DFO). The presented study reassesses the impact of DFO as a virulence factor of *E. amylovora* during its epiphytic phase on the apple flower. When inoculated in semisterile Golden Delicious flowers no difference in replication and induction of calyx necrosis could be observed between *E. amylovora* CFBP1430 siderophore synthesis (DfoA) or uptake (FoxR receptor) mutants and the parental strain. In addition, mutant strains only weakly induced a *foxR* promoter-*gfpmut2* reporter construct in the flowers. When analyzing the replication of the receptor mutant in apple flowers harboring an established microbiome, either naturally, in case of orchard flowers, or by pre-inoculation of semisterile greenhouse flowers, it became evident that the mutant strain had a significantly reduced replication compared to the parental strain. The results suggest that apple flowers *per se* are not an iron-limiting environment for *E. amylovora* and that DFO is an important competition factor for the pathogen in precolonized flowers.

IMPORTANCE Desferrioxamine is a siderophore produced by the fire blight pathogen *E. amylovora* under iron-limited conditions. In the present study, no or only weak induction of an iron-regulated promoter-GFP reporter was observed on semisterile apple flowers, and siderophore synthesis or uptake (receptor) mutants exhibited colonization of the flower and necrosis induction at parental levels. Reduced replication of the receptor mutant was observed when the flowers were precolonized by microorganisms. The results indicate that apple flowers are an iron-limited environment for *E. amylovora* only if precolonization with microorganisms leads to iron competition. This is an important insight for the timing of biocontrol treatments.

KEYWORDS *Erwinia amylovora*, desferrioxamine, siderophore mutants, apple flowers, iron deficiency, replication, secondary colonization

Even though iron is the fourth most abundant element on Earth, under oxidative conditions its bioavailability is limited to microorganisms. This limitation is due to the reduction of Fe²⁺ to the insoluble Fe³⁺ state. As iron is an essential cofactor in reactions such as DNA replication or protection against oxygenated radicals, many microorganisms have evolved high-affinity systems to acquire iron, so-called siderophores. Siderophores are

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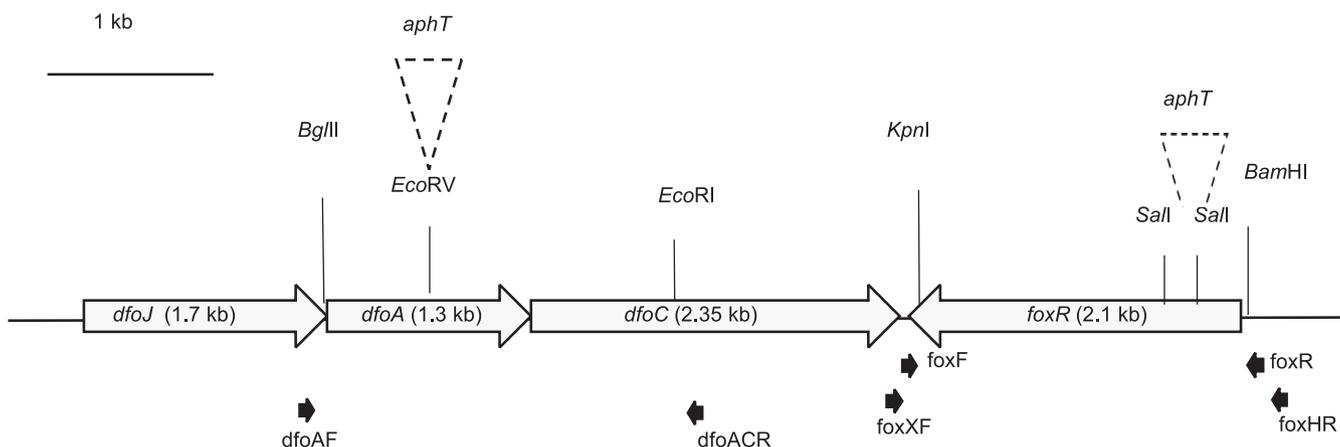


FIG 1 Schematic representation of the ferrioxamine cluster in *E. amylovora* CFBP1430. Large gray arrows indicate coding sequences. Small black arrows show the position and direction of primers used for site-directed mutagenesis of the *dfoA* and *foxR* genes. A kanamycin resistance cassette (*aphT*) was inserted into the *EcoRV* cutting site of the *dfoA* gene, resulting in the desferrioxamine synthesis mutant EAdfoA. Replacing the 203 pb *Sall* fragment of the *foxR* gene by *aphT* resulted in the ferrioxamine receptor mutant EAfoxR.

molecules with low molecular masses (200 - 2000 Da) that are produced by bacteria when the intracellular iron concentration is low (1, 2). Siderophores chelate Fe^{3+} with a very high affinity and the chelated complexes are recognized by highly selective outer-membrane TonB-dependent receptors on the cell surface of Gram-negative bacteria. The receptors bind the Fe^{3+} siderophore complex, which is then actively transported across the membrane through an energy-dependent system into the cytosol, where the iron is released (3). The regulation of iron metabolism in bacteria is mediated by the ferric-uptake regulator protein (Fur). Fur represses transcription of iron-uptake associated genes upon interaction with its corepressor Fe^{2+} by binding the consensus sequence known as "Fur box" within Fur-regulated promoters (4, 5). The fire blight pathogen *Erwinia amylovora* is considered to be the most important threat to pome fruit production (e.g., apple, pear, and quince) globally. The pathogen is vectored by flower-foraging insects, mainly bees (6, 7). It infects the host tissue through the nectarhodes after an epiphytic phase during which it replicates on the flower stigma. *E. amylovora* produces hydroxamate-type desferrioxamine siderophores (DFOs), mostly DFO E (nocardamine) (8, 9). DFO E is encoded by the *dfoJAC* gene cluster, the specific receptor for the passage of the ferric complex across the outer membrane by *foxR*. The *foxR* gene is located on the complementary strand directly downstream of *dfoC* (Fig. 1). Both *foxR* and the *dfoJAC* cluster are regulated by Fur via their Fur box containing promoters (10, 11).

For *E. amylovora* CFBP1430 a dual function of DFO has been proposed. The siderophore plays a role in cell protection against the host oxidative burst elicited during infection and is critical for its iron acquisition (12). Transposon mutants either defective in the DFO biosynthetic pathway (*dfoA* mutant) or uptake of the ferric complex (*foxR* mutant) have been studied. Whereas a *dfoA* mutation disrupts the synthesis of the siderophore, a mutation in the FoxR receptor leads to an accumulation of the siderophore in the external medium due to the lack of reuptake (10, 12). When tested on apple flowers the *dfoA* and *foxR* mutants revealed a reduction in their growth (2 orders of magnitude) and in their ability to initiate necrotic symptoms compared to the wild-type strain (12).

Studies on a pyoverdine siderophore negative mutant of *Pseudomonas orientalis* F9 which inhibited growth of *E. amylovora* as successful as the parental strain (13) prompted us to reassess the impact of DFO as a virulence factor of *E. amylovora* during the epiphytic phase in the apple flower. To this end siderophore synthesis (*dfoA*) and uptake (*foxR*) mutant strains were tested on various Golden Delicious (GD) flowers for their growth rates, calyx necrosis, and induction of a *foxR* promoter-*gfpmut2* reporter construct.

In semisterile GD flowers siderophore mutants have no disadvantage in terms of

TABLE 1 Strains used in this study

Strain	Genotype and/or phenotype	Reference
<i>Escherichia coli</i> DH5 α	<i>endA1 hsdR17</i> ($r_K^- m_K^+$) <i>supE44 thi-1 recA1 gyrArelA1</i> Δ (<i>lacZYA-argF</i>)U169 (80 <i>lacZ</i> Δ M15)	41
<i>E. coli</i> S17-1 λ	<i>pir</i> ⁺ <i>tra</i> ⁺	32
<i>Erwinia amylovora</i> CFBP1430	Isolated in 1972 from a <i>Crataegus</i> sp.	42
EA ^S : <i>E. amylovora</i> CFBP1430 Sm ^R	Spontaneous streptomycin resistant mutant of <i>Erwinia amylovora</i> CFBP1430	This study
EA ^S gfp: <i>E. amylovora</i> CFBP1430 Sm ^R , (pEA <i>foxRgfp</i> mut2) Amp ^R	Spontaneous streptomycin resistant mutant of <i>Erwinia amylovora</i> CFBP1430, carrying reporter plasmid <i>pfoxR</i> promoter- <i>gfp</i> mut2	This study
EAdfoA: <i>E. amylovora</i> CFBP1430EAdfoA::aphT Sm ^R , Kan ^R	Siderophore negative mutant of EA ^S , CAS negative	This study
EAdfoAgfp: <i>E. amylovora</i> CFBP1430EAdfoA::aphT (<i>pfoxRgfp</i> mut2), Amp ^R , Sm ^R , Kan ^R	Siderophore negative mutant of EA ^S , carrying reporter plasmid <i>pfoxR</i> promoter- <i>gfp</i> mut2	This study
EAfoxR: <i>E. amylovora</i> CFBP1430 <i>foxR</i> ::aphT, Sm ^R , Kan ^R	Siderophore receptor mutant of EA ^S , CAS halo oversized	This study
EAfoxRco: <i>E. amylovora</i> CFBP1430EAfoxR::aphT, (pKS <i>foxR</i>), Amp ^R , Sm ^R , Kan ^R	Siderophore receptor mutant of EA ^S , partially complemented with plasmid pKS <i>foxR</i>	This study
EAfoxRgfp: <i>E. amylovora</i> CFBP1430 <i>foxR</i> ::aphT, (<i>pfoxRgfp</i> mut2), Amp ^R , Sm ^R , Kan ^R	Siderophore receptor mutant of EA ^S carrying reporter plasmid <i>pfoxR</i> promoter- <i>gfp</i> mut2	This study
PW: <i>Pantoea vagans</i> C9-1W	<i>Pantoea vagans</i> C9-1 variant that Lacks the 530-kb megaplasmid pPag3. pPag3 encodes among other things (e.g., carbon utilization, thiamin, carotenoids) the desferrioxamine synthesis genes.	43
PWgfp: <i>P. vagans</i> C9-1W (<i>pfoxRgfp</i> mut2), Amp ^R	<i>Pantoea vagans</i> C9-1W carrying reporter plasmid <i>foxR</i> promoter- <i>gfp</i> mut2	This study

replication and necrosis induction compared to the parental strain, nor does *E. amylovora* CFBP1430 induce *foxR* when colonizing the stigma. Significant growth deficiency of a receptor mutant was observed in pre-colonized flowers, either by a naturally evolved microbiome in the orchard or by pre-incubation of a competitor on semisterile greenhouse flowers. It has been shown that precolonization of semi sterile leaves can drastically decrease the reproductive success of secondary colonizers (14). The present study shows how the priority effects of colonization alter the context-specific dependence of a pathogenicity factor during flower colonization.

The results indicate that the DFO system of *E. amylovora* is required when the pathogen performs as a secondary flower colonizer.

RESULTS

Siderophore synthesis and growth of parental and mutant strains. Parental strain EA^S (*E. amylovora* CFBP1430 Sm^R, Table 1), corresponding desferrioxamine synthesis mutant *E. amylovora* EAdfoA, ferrioxamine receptor mutant EAfoxR and complemented mutant EAfoxRco were analyzed on CAS detection agar for siderophore synthesis (Fig. 1, Fig. 2). As expected, EAdfoA was not able to produce an orange halo indicative for siderophore synthesis. The receptor mutant EAfoxR produced a larger halo than the parental strain EA^S due to its inability to take up the siderophore-iron complex and hence inability to repress the expression of siderophore production. After complementation of EAfoxR with the plasmid pEA*foxR* (EAfoxRco) the halo size was reduced.

While growing in LB medium, all strains grew similarly with the exception of the complemented strain EAfoxRco which showed slightly impaired growth (Fig. 3A). When inoculated in iron-limited KB medium, EAfoxR showed a significant growth reduction compared to the parental strain, which was partially restored in the complemented strain EAfoxRco (Fig. 3B).

Induction of flower necrosis. The ability of the strains to induce necrosis in apple flowers was tested on detached GD flowers. The infection grades (Fig. 4) and the resulting severity grades of parental strain EA^S (71.5), siderophore synthesis mutant EAdfoA (73.5), and receptor mutant EAfoxR (71.5) revealed no difference in the induction of flower necrosis.

Bacterial densities on the stigma and hypanthium. In the established assay for induction of flower necrosis as performed above, the inoculum is directly applied onto

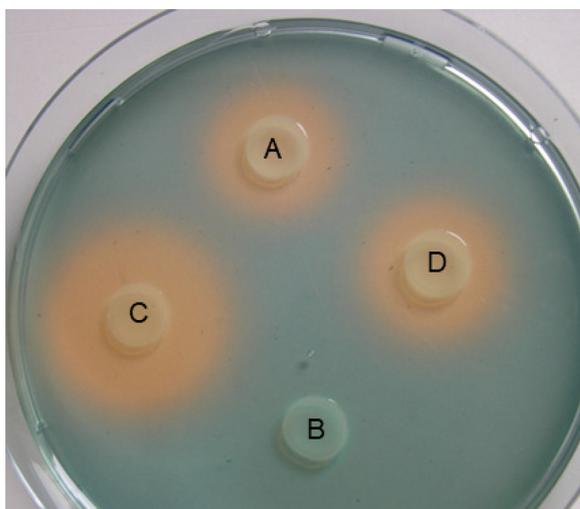


FIG 2 Siderophore indication CAS-agar with *E. amylovora* CFBP1430 parental strain EA^S (A), desferrioxamine synthesis mutant EAdfoA (B), ferrioxamine receptor mutant EAfoxR (C) and complemented receptor mutant EAfoxRco (D). The orange halos are indicative for siderophore production.

the hypanthium of the flowers. Therefore, the assay omitted the important step of the replication of the pathogen on the stigmata. To assess the impact of the mutations on the replication, stigmata of detached GD flowers were dipped into bacterial suspensions of EA^S, EAdfoA, EAfoxR, and the complemented receptor mutant EAfoxRco. Parental and mutant strains replicated equally on stigmata and only the complemented EAfoxRco strain revealed a slightly reduced CFU on the hypanthium (Fig. 5).

Bacterial densities and necrosis induction in flowers of GD trees. Three flowering GD trees each were sprayed with bacterial suspensions of EA^S, EAdfoA and EAfoxR. In accordance with the previous results, there were no differences in the recovered CFU. The same was true for the induction of flower necrosis (Fig. 6A and B).

FoxR promoter activity on apple flower stigmata. None of the *in vivo* assays performed revealed a difference between *E. amylovora* parental and siderophore mutant strains regarding their replication on GD flowers and induction of necrosis. To verify the need for the siderophore system during the epiphytic phase of the fire blight pathogen, the Fur regulated *foxR* promoter was fused to the *gfpmut2* encoding gene and the resulting reporter plasmid (*pfoxRgfpmut2*) transformed into the parental and both mutant strains resulting in EA^Sgfp, EAdfoAgfp, and EAfoxRgfp. To test the functionality of the reporter-construct EAfoxRgfp was inoculated in iron deficient KB medium. In addition to the *E. amylovora* strains, the white variant of the well-studied *E. amylovora* antagonist *P. vagans* C9-1, *P. vagans* C9-1W (PW), was included in the assay as fluorescent positive-

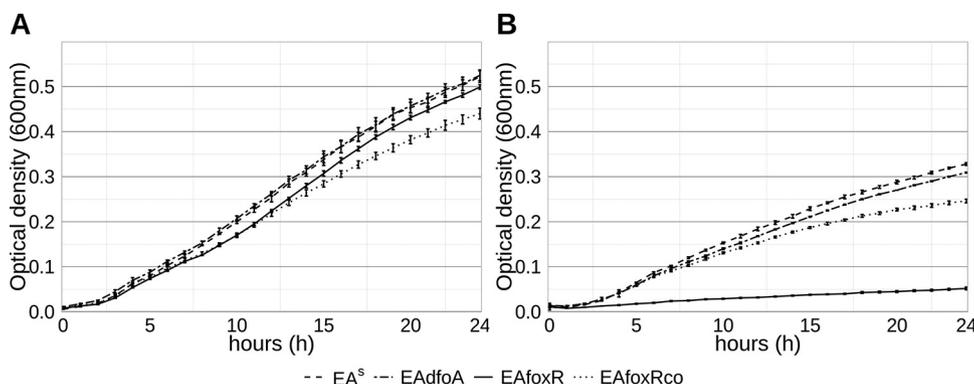


FIG 3 LB (A) and KB (B) growth curves of *E. amylovora* CFBP1430 strains EA^S (solid), desferrioxamine synthesis mutant EAdfoA (dotted), ferrioxamine receptor mutant EAfoxR (dashed) and complemented mutant EAfoxRco (dotdash). Error bars represent standard deviations.

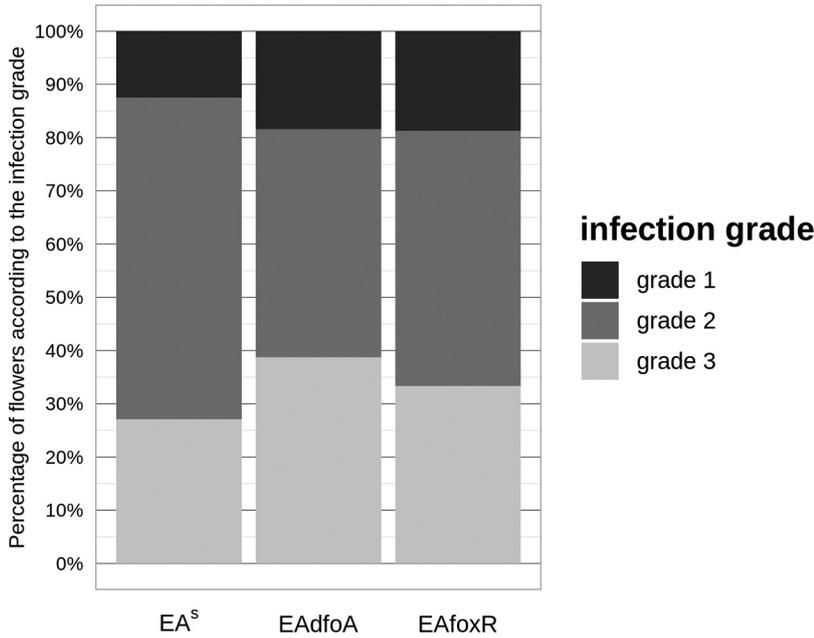


FIG 4 Percentage fraction of flowers in each infection grade. Each column represents 48 evaluated flowers from two independent detached flower assay experiments. Flowers were inoculated onto the hypanthium with *E. amylovora* CFBP1430 strains EA^S, desferrioxamine synthesis mutant EAdfoA, and ferrioxamine receptor mutant EAfoxR. Evaluation of the flowers infection grade was performed after 4 days of incubation at 26°C according to the following scale: grade 1: calyx green; grade 2: calyx necrotic; grade 3: calyx and pedicel necrotic.

control PWgfp. PW is deficient of the megaplasmid pPag3, which carries the desferrioxamine siderophore encoding genes in this strain (Table 1). GFP measurements showed an induction of the reporter construct in both strains (Fig. S1).

When GFP measurements were performed *in planta*, none of the *Erwinia* strains tested showed a strong induction of the *foxR* promoter reporter (Fig. 7). The percentage of *gfp* induced cells was between 5% (EAfoxRgfp) to 10% (EAdfoAgfp), (Table 2). The parental strain showed no induction (0.4%). This is in contrast to control strain PWgfp, where 85% of the cells were GFP positive (Fig. S2), confirming the assay to be able to detect high GFP levels, if the reporter is induced *in planta*.

Bacterial densities on the stigmata of greenhouse or orchard-grown GD flowers.

In contrast to the previous study (12) apple flowers from the above performed experiments originated from a closed compartment and were not exposed to nectar foraging insects, wind or rain. Thus, the observed difference is likely due to the colonization of

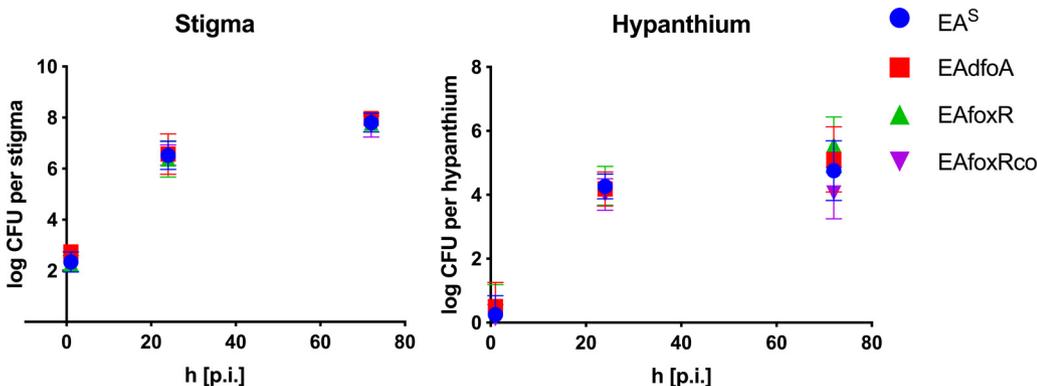


FIG 5 CFU of parental strain *E. amylovora* CFBP1430 EA^S, corresponding desferrioxamine synthesis mutant (EAdfoA), ferrioxamine receptor mutant (EAfoxR), and complemented ferrioxamine receptor mutant (EAfoxRco) 0 h, 24 h and 72 h p.i. on stigma and hypanthium of detached GD flowers.

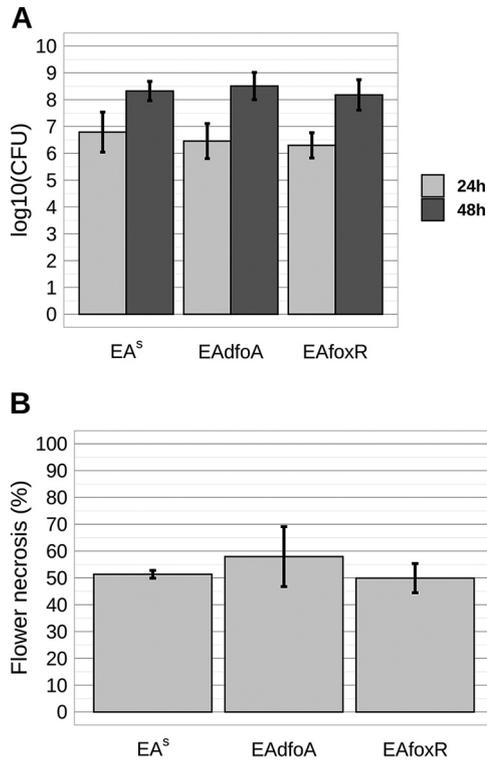


FIG 6 CFU and necrosis of GD flower calyxes after spray inoculation of flowering GD trees with *E. amylovora* CFBP1430 strains EA^S, EAdfoA, and EAfoxR. (A) CFU were determined for 10 flowers per strain 24 h and 48 h p.i. Error bars represent the standard deviation from the mean. (B) Disease symptoms of flowers rated 6 days after inoculation. Error bars represent the standard deviation of three trees. In total 413, 474 and 333 flowers were evaluated for EA^S, EAdfoA and EAfoxR, respectively.

the flowers with a microbiome that develops in an orchard environment but not in a closed compartment. To test this hypothesis, semisterile GD flowers from the greenhouse and open (microbiome bearing) flowers from the orchard-P23 were inoculated with EA^S or the EAfoxR receptor mutant. In semisterile flowers from the greenhouse both strains replicated equally (Fig. 8A). In accordance with the results of the previous study (12) the EAfoxR mutant revealed a reduced CFU in the orchard-P23 flowers compared to the parental strain (Fig. 8B). The complemented receptor mutant (EAfoxRco)

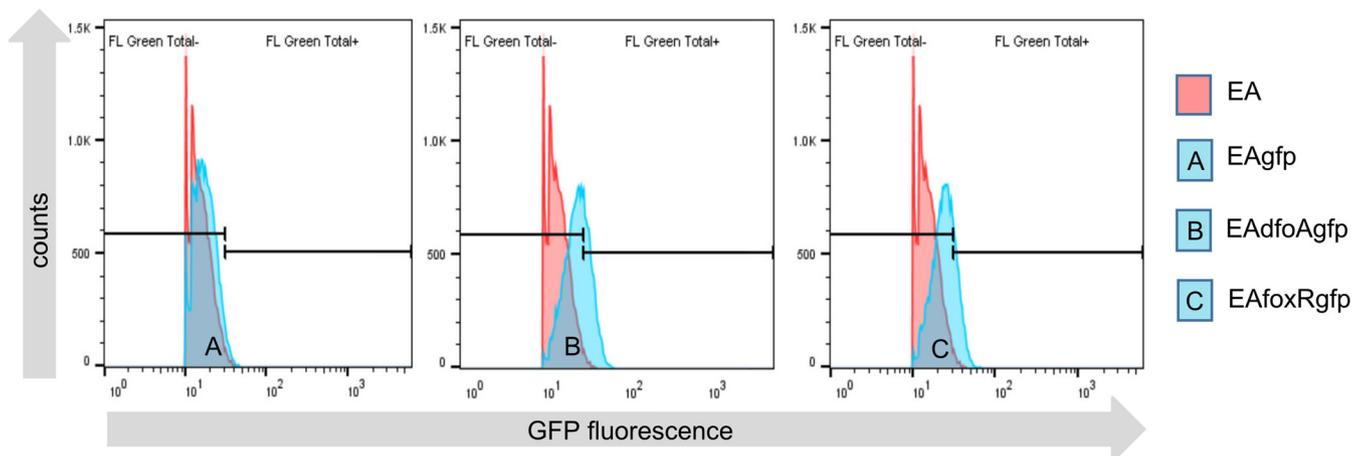


TABLE 2 Single cell GFP fluorescence of reisolated reporter strains 48 h p.i. from GD apple stigmata

Strain	Total count	Count GFP positive	Count GFP negative	% GFP positive
EA	21083	71	21012	0.3
EAgfp	22594	92	22502	0.4
EAdfoAgfp	21552	2214	19338	10.3
EAfoxRgfp	20149	962	19187	4.8

multiplied as efficiently as the parental strain in microbiome containing orchard-P23 flowers (Fig. 8C, Fig. S3). When EA^S and EAfoxR were inoculated in freshly opened, semisterile balloon flowers from the orchard-P23, both strains multiplied equally well (Fig. 8D).

To verify the CFU data gained by classical plating, an EA specific qPCR was performed on each of the flower samples from open- (EA^S, EAfoxR, EAfoxRco) and balloon orchard-P23 flowers (EA^S and EAfoxR, Fig. 9), confirming the replication deficiency of the receptor mutant in open orchard flowers.

Bacterial densities on the stigmata of pre-inoculated greenhouse GD flowers.

To verify the impact of flower colonizers on the replication of receptor mutant and to exclude abiotic factors and changes in the nutrient composition of the open orchard flowers to be the main cause, freshly opened GD flowers from the greenhouse were pre-inoculated with PW as a competitor. Consistent with the results from open orchard-P23 GD flowers, EA^S was significantly better at colonizing the stigma than EAfoxR (Fig. 10). Performing the *amsC* qPCR using DNA templates consisting of 10 pooled samples confirmed the CFU data obtained (Fig. S4). The complemented strain EAfoxRco reached the parental CFU level in the pre-inoculated flowers (Fig. S5).

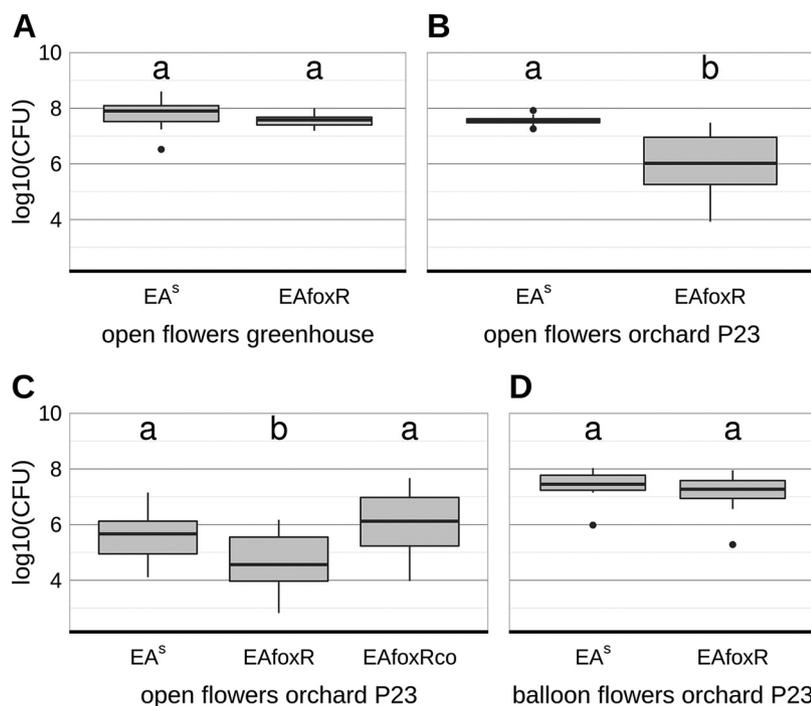


FIG 8 CFU of reisolated *E. amylovora* CFBP1430 strains 48 h p.i. (A) parental strain EA^S (*n* = 9) and mutant EAfoxR (*n* = 20) from semisterile GD flowers, greenhouse; (B) parental strain EA^S (*n* = 10) and mutant EAfoxR (*n* = 20) from open GD flowers, orchard-P23; (C) parental strain EA^S (*n* = 20), mutant EAfoxR (*n* = 20) and complemented mutant strain EAfoxRco (*n* = 19) from open GD flowers, orchard-P23; (D) parental strain EA^S (*n* = 10) and mutant EAfoxR (*n* = 9) from GD balloon flowers, orchard-P23. Error bars represent the standard deviation of the mean. Significant differences between treatments are marked with different letters (*P*-value < 0.05, one-way ANOVA, Tukey's multiple-comparison test).

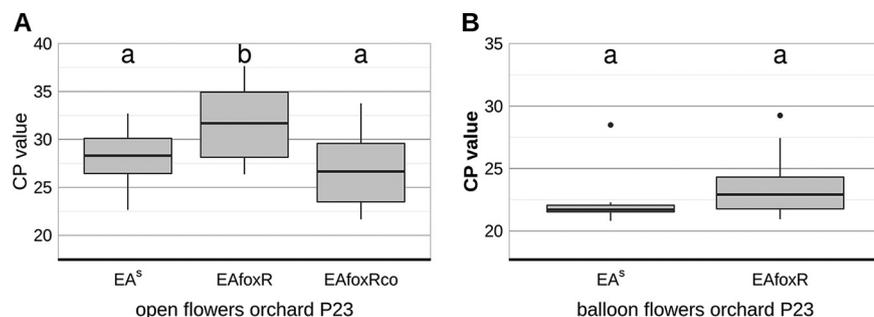


FIG 9 CP values of *E. amylovora* CFBP1430 strains. (A) EA^s ($n = 20$), corresponding mutant EAfoxR ($n = 20$) and complemented mutant strain EAfoxRco ($n = 19$) reisolated from open GD flowers, orchard-P23 after 48 h p.i.; (B) EA^s ($n = 10$) and mutant EAfoxR ($n = 10$) reisolated from GD balloon flowers, orchard-P23, 48 h p.i. The bacterial DNA was extracted from each infected flower and qPCR performed with an *amsC* (*amylovoran* synthesis) specific probe. Error bars represent the standard deviation of the mean. Significant differences between treatments are marked with different letters (P -value < 0.05, one-way ANOVA, Tukey's multiple-comparison test).

DISCUSSION

The synthesis and regulation of siderophores, Fe³⁺ chelators (15), have been intensively studied in human-pathogenic bacteria, e.g., *Yersinia* and *Salmonella enterica*, demonstrating their role in mediating pathogen multiplication and development of virulence (16–18). The role of siderophores in plant microbial pathogenesis is less well studied. From *Dickeya dadantii* 3937, the causal agent of “soft rot,” it is known that the synthesis of siderophores is required for the systemic progression of maceration symptoms in the host (19) and *Pseudomonas syringae* pv. *tabaci* 6605 requires the siderophore pyoverdine for full virulence in tobacco (20). *E. amylovora*, the causal agent of fire blight produces desferrioxamine DFO siderophores, mainly DFO-E (8, 9). *E. amylovora* mutants defective in siderophore synthesis or uptake exhibited a reduced ability to colonize floral tissues and to cause necrosis, indicating that DFO is a virulence factor during the onset of infection (12). The presented study focuses on the DFO system and its impact on *E. amylovora* during the epiphytic state of the pathogen on the apple flower. To this end, a *dfoA* (EAdfoA) siderophore synthesis and a *foxR* (EAfoxR) receptor mutant were constructed. Neither the EAdfoA nor the EAfoxR mutant revealed a decrease in replication or in necrosis induction when applied onto GD flowers from a closed greenhouse. Additionally, a fluorescence whole-cell bio-reporter under the control of the Fur regulated *foxR* promoter was not induced in the parental strain and only weakly (5% to 10%) in the mutant strains when incubated on greenhouse-grown stigmata (Fig. 7). In contrast, in iron-limited KB medium, 93% (Fig. S1) of the receptor mutant cells were classified as GFP-positive.

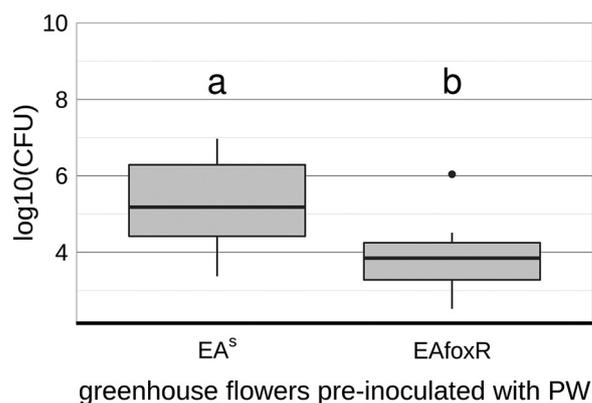


FIG 10 CFU of *E. amylovora* CFBP1430 strains EA^s ($n = 20$) and corresponding mutant EAfoxR ($n = 20$) 48 h p.i. onto PW pre-inoculated semisterile GD flowers from the greenhouse. Error bars represent the standard deviation of the mean. Significant differences between treatments are marked with different letters (P -value < 0.05, one-way ANOVA, Tukey's multiple-comparison test).

These results imply that semisterile GD flowers do not represent an iron-limiting environment for *E. amylovora* and therefore, the lack of siderophore production or uptake is not a disadvantage in replication and necrosis induction for mutant strains. This is in contrast to *P. fluorescens* strain A506, an antagonist of *E. amylovora*, commercialized as BlightBan. An ice nucleation protein-based promoter reporter assay demonstrated that pyoverdine siderophore genes are upregulated on apple and pear flowers in growth conditions where blossoms were protected from rain and insect visitations, which indicates that those blossoms represented an iron-limited environment to *P. fluorescens* A506 (21).

The *E. amylovora* CFBP 1430 genome contains only remnants of the siderophore receptors ferrichrome (FhuA) and aerobactin (lutA) (22). The inactivation of such a potential source of Fe³⁺ acquisition possibly indicates that *E. amylovora* only requires siderophores for replication at some steps of its infection cycle, not necessarily the flower. It might be that *E. amylovora* compensates its iron need by additional systems present on its chromosome, e.g., SitABCD/YfeA-E that also can take up Fe²⁺ or the iron specific EfeUOB system (23–27).

The flowers used in these experiments originated from an enclosed facility and thus, were semisterile. Studies on the expression of *foxR* in inoculated apple leaves (28) already revealed differential expression of the gene depending on localization and bacterial density. Therefore, it is possible that the siderophore is critical for *E. amylovora* replication in the flower only if the pathogen competes with additional apple blossom colonizers. To test this hypothesis, open GD flowers from the orchard P23 were inoculated with the EA^S and the EAfoxR strain. The results confirmed a decreased ability of EAfoxR to replicate on precolonized flowers, which was reversed when the mutant strain was complemented (Fig. 8C and 9A). To verify that EAfoxR growth inhibition is due to competitors, freshly opened greenhouse-grown GD flowers were pre-inoculated with the strain PW. Similar to flowers grown in a natural environment, EAfoxR, in contrast to the complemented mutant, could not replicate to the same density as the parental strain.

The here presented results imply that iron is readily bioavailable to *E. amylovora* during its epiphytic phase on non-precolonized flowers. In already colonized flowers, either with a complex microbiome or a single competitor, EAfoxR growth decreased to an extent that indicates the siderophore system as an important competitive factor in flower colonization. This underscores the importance of an initial flower colonization by epiphytes in pest management of *E. amylovora*.

MATERIALS AND METHODS

Cultivation of bacteria. For apple flower inoculation and CFU determination after reisolation from infected flowers *E. amylovora* strains were grown at 26°C in Tryptic Soy broth (TSB, Oxoid) or on TSB plates (addition of 15 g liter⁻¹ agar [Roth]). *E. coli* strains were cultivated at 37°C in Lysogeny broth (LB, Carl Roth) or on LB plates (addition of 15 g liter⁻¹ agar). For growth curve experiments *E. amylovora* strains were grown in King's B medium (29) and in Lysogeny broth. Microorganisms used in the study are listed in Table 1. Where appropriate, medium was supplemented with kanamycin 40 mg liter⁻¹, streptomycin 100 mg liter⁻¹, rifampicin 100 mg liter⁻¹, ampicillin 100 mg liter⁻¹ or cycloheximide 100 mg liter⁻¹, respectively.

Construction of mutant strains. To inactivate the *foxR* gene, a part of the gene was amplified using primers FoxF (5'-AAGCTAAACCGCGATAAGTATAG-3') and EAfoxR (5'-TCGTAACCGACGGTAGCCATC-3') resulting in a 2.1 kb fragment harboring a BamHI and KpnI cutting site at the 5' and 3' prime end, respectively. The amplified fragment was digested with BamHI/KpnI and ligated into similarly digested plasmid pBluescript (pKS). Subsequently, the resulting vector was digested with Sall, which cuts twice within the open reading frame of the *foxR* gene, deleting 203 bp (Fig. 1). The excised region was replaced with a HindIII cut kanamycin cassette from plasmid pSB315 (*aphT*), which lacks a transcriptional terminator (30). Using primers M13F (5'-CAGGAAACAGCTATGAC-3') and M13R (5'-TGTAACGACGGCCAGT-3') the *foxR*-kanamycin cassette was amplified, digested with BamHI/KpnI and ligated into the similarly digested suicide vector pKAS32 (31). The resulting construct, pSVEAfoxR was transformed into *E. coli* S17-1 (32, 33) and mobilized into a spontaneous streptomycin resistant mutant of *E. amylovora* CFBP1430 (EA^S). Plasmid pKAS32 contains the *rpsL* gene encoding the S12 protein of the ribosomes, which is the target of streptomycin. Thereby insertion of the suicide vector into the chromosome results in a streptomycin sensitive phenotype of the formerly resistant strain, which counter selects double crossing over events. The receptor mutant EA^S*foxR*::*aphT* (EAfoxR) was selected on TSB agar plates containing kanamycin and streptomycin. The insertion was confirmed by PCR and subsequent sequencing of the PCR product.

For complementation, the *foxR* gene and its native promoter were amplified using primers FoxHR (5'-AAAAAAGCTTAACGCTCTCTCTCTGGG-3') and FoxXF (5'-GGGGGICTAGAAAGCTAAACCGCGAT

AAGTATAG-3') which introduce a XbaI and HindIII cutting site, respectively (underlined). The resulting 2.6 kb PCR product was XbaI/HindIII digested and ligated into the similarly digested pKS vector. The resulting plasmid pKS foxR was electroporated into EA foxR to generate the complemented mutant EA foxRco .

To inactivate the *dfoA* gene, a 2.3 kb fragment of the gene was amplified using primers dfoA2661F (5'-CTGCTGCACAACGTGCTGCT-3') and dfoA4940R (5'-GTGCATTACGGCCGTGAACA-3') harboring an EcoRI and a BglII cutting site at the 5' and 3' end of the PCR product, respectively. The PCR product was EcoRI/BglII digested and ligated into the similarly digested suicide vector pKAS32 and transformed into *E. coli* S17-1. The plasmid was EcoRV digested which cuts within the *dfoA* gene and the HindIII cut kanamycin cassette from plasmid pSB315 was inserted. The resulting plasmid pSV dfoA was subsequently mobilized into EA⁵. EA⁵*dfoA::aphT* (EA dfoA) mutants were selected on KB agar plates containing kanamycin and streptomycin and confirmed by PCR.

Construction of the *foxR* promoter-GFPmut2 construct. To construct a *foxR* promoter-GFP reporter construct, 365 bp upstream up the *foxR* gene (including the upstream regulatory Fur sequences) were amplified using primers P fox_XF (5'GGGGGCTAGAAAGCTAAACCACGCGATAAGTATAG3') and P fox_PR (5'GGGGGCTGCAGTTTATCCCTTACAAAGATTA3') with designed restriction sites XbaI and PstI (underlined). The resulting PCR product was XbaI/PstI digested and ligated into similarly digested plasmid pM965 (34) which places the GFPmut2 encoding gene in pM965 under the control of the *foxR* promoter. The resulting plasmid p foxRgfpmut2 was electroporated into the *E. amylovora* parental and mutant strains and *P. vagans* C9-1W.

Siderophore indicator test. Siderophore production was tested on Chrome azurol S (CAS) agar (35). Freshly grown colonies of EA⁵, EA dfoA , EA foxR , and EA foxRco grown on TSB agar plates with the appropriate antibiotics overnight were resuspended in 1 × PBS buffer (NaCl 8 g liter⁻¹, KCl 0.2 g liter⁻¹, Na₂HPO₄·12H₂O 2.9 g liter⁻¹, KH₂PO₄ 0.2 g liter⁻¹, pH 7.2) to an OD_{600nm} = 1. Five μL of the resuspended cultures were applied onto the indicator agar. Formation of an orange halo around applied bacteria was indicative for siderophore production.

Growth curves. Bacterial growth was monitored using a Bioscreen C (Oy Growth Curves Ab Ltd., Helsinki). To study the effect of iron deficiency on EA⁵ and corresponding mutants, growth was observed in LB and iron-limited KB medium. Seven hundred μL of the respective media were inoculated with 3 μL of an overnight culture and loaded in three replicates (200 μL per well) on a Bioscreen C honeycomb plate. For each replicate 200 μL of the corresponding medium were used as a negative control. Measurement of optical density at 600 nm (OD_{600nm}) was performed every hour for 24 h at 26°C with 10 sec of shaking prior to each measurement. Two experiments were repeated independently.

Inoculation of flowering apple trees in the greenhouse. 3-year-old GD trees were potted and kept in a quarantine greenhouse (closed compartment) until flowering. Selected strains were applied on each single flower using a hand sprayer (200 μl per hub) and an OD_{600nm} = 1 bacterial suspension (see next paragraph) that was 10⁻²-fold diluted in PBS buffer. Three apple trees per strain were placed in a greenhouse cabin with 60% humidity after spray inoculation. The temperature was set to 18°C (10 h) at night and 24°C (10 h) at day and 2 h ramp time. After 6 days, flower necrosis symptoms were evaluated, indicated by a darkening of tissue of the flower calyx (brownish to black: necrosis positive, green: necrosis negative).

To estimate the CFU of bacteria in spray inoculated flowers, flower petals, pedestals, and stems were removed. The remains of the apple blossom were shaken in 1 mL PBS buffer for 30 min at 1400 rpm and then vortexed for 30 sec. A serial dilution of the resulting supernatants was performed up to a 10⁻⁷-fold dilution. Three μl of each dilution step from each sample were transferred onto TSB agar plates containing appropriate antibiotics using a 96-replicate plate (Fig. S6). As a control, defined EA suspensions were equally diluted and spotted alongside the samples. CFU counts were performed after 24 h under a binocular microscope. For resulting supernatants that led to no cultivated colonies on the TSB agar plates, the minimal detection limit of one colony was calculated.

Detached flower assay, visual grading. The detached flower assay (36) was performed as previously described (37). Freshly opened flowers of 2-year-old potted GD trees were used. In autoclaved Eppendorf racks every second well of every second row was filled with two mL autoclaved tap water and sealed with office tape. Holes were punched through the tape using sterile pipette tips and finally the stems of detached flowers were inserted into the holes.

All used EA strains were grown overnight on TSB agar plates. Bacterial biomass was resuspended in PBS buffer to an OD_{600nm} = 1. Twenty μL of each bacterial suspension was added to 980 μL PBS buffer. Twenty μL of the dilution were then directly pipetted onto the hypanthium of individual flowers. Control treatments were performed using PBS buffer only.

For the three strains EA⁵, EA dfoA and EA foxR , a total of 2 × 24 flowers were tested in two independent experiments. After inoculation of the flowers, the Eppendorf racks were transferred in a storage box laid out with water-soaked paper towels to ensure high relative humidity. Incubation was performed at 26°C for four to 5 days. After incubation each flower was visually graded based on the following scale: grade 1: calyx green; grade 2: calyx necrotic (brownish), pedicel green; grade 3: calyx and pedicel necrotic (Fig. S7). The results of both experiments were added up and the severity grade of the infection was defined as previously described (38).

Growth on stigma and hypanthium. To estimate the CFU of parental strain EA⁵ and mutant strains on the stigma and hypanthium of detached flowers, the OD_{600nm} = 1 (see above) bacterial suspensions were 10⁻⁴-fold diluted and 1.5 mL reaction tubes filled up to the rim with these suspensions. The petals and stamens of GD flowers were removed with sterile scissors and the exposed stigmata were inoculated by carefully dipping them into the bacterial suspension. Immediately after drying of the bacterial suspension, the CFU of bacteria on stigma and hypanthium of single flowers were determined

(time = 0). Stigmata of each flower were cut with sterile scissors and collected in a 1.5 mL reaction tube filled with 200 μ L PBS buffer. The remains of the flowers were similarly treated. For CFU determination after prolonged incubation (24 h and 72 h), flower parts were resuspended in 1 mL PBS buffer. The CFU determination was performed as described above in section "Inoculation of flowering apple trees in the greenhouse."

FCM (Flow cytometer) analysis. For *in vitro* measurements of GFP expressing bacteria, a TSB overnight culture of the strains containing the appropriate antibiotics was diluted 1:100 and cultivated (240 rpm) at 26°C for 48 h. Afterwards, 1 mL of the bacterial suspension was centrifuged (30 sec, max. speed, Eppendorf MiniSpin) and the pellet washed in sterile filtered PBS.

After an additional centrifugation step the pellet was resuspended in 100 μ L PBS-buffered 4% paraformaldehyde and stored at 4°C in the dark before processing.

For GFP measurement *in planta*, bacteria were cultivated overnight and bacterial suspensions adjusted to OD_{600nm} = 1 in PBS buffer were diluted 10⁻⁵-fold. The stamens of detached flowers were removed with sterile scissors and the exposed stigmata inoculated with 2 μ L of the diluted suspensions. Inoculated flower stigmata were incubated at 26°C for 24 and 48 h. After incubation the stigmata of each flower were cut with sterile scissors and collected in a 1.5 mL reaction tube containing 100 μ L sterile filtered PBS. Samples were shaken at RT for 30 min at 1400 rpm. Subsequently, the tubes were vortexed vigorously for 2 min. Ninety μ L of the suspension were carefully transferred by slowly pipetting into 1.5 mL reaction tubes with 90 μ L PBS-buffered 4% paraformaldehyde (Alfa Aesar). Samples were kept at 4°C in the dark for up to 24 or 48 h before the FCM analysis was performed.

Flow cytometric measurements were performed with the CytoSense benchtop FCM (Cytobuoy, Netherlands). The FCM is equipped with a 125 mV laser at a wavelength of 488 nm. The green fluorescence of GFP was detected in a range from 509 to 540 nm. Each sample was diluted with sterile PBS until a particle count between 500 and 5'000 particles per second was present during measurement (for ca. 20000 cells per flower sample). To distinguish between impurities and cells of interest a gate (subset of all measured events) was established based on forward scatter (FWS) and sideways scatter (SWS) signals. As a control for gate selection Image In Flow (IFF) pictures were taken into account. Based on the gate the fluorescence intensity of the GFP (FI green signal integral) was used to differentiate between GFP positive and GFP negative cells.

Competition studies using GD orchard or greenhouse flowers. GD flowers were collected in the apple orchard-P23 at the Agroscope Research Station in Wädenswil, Switzerland (GPS: 47°13'18.1" N, 8°40'38.9" E). To ensure sufficient time for the development of a microbiome in the flowers, the collected flowers had their petals open and partially brownish anthers (Fig. S8). Flowers collected in the balloon state and thus devoid of a microbiome were incubated at 26°C for several hours to trigger petal opening and allow subsequent inoculation. For pre-inoculation of flowers with the competitor *P. vagans* C9-1W (PW), GD flowers from the greenhouse were used (Fig. S8).

All inoculations for orchard flowers were performed by pipetting 2 μ L of a 10⁻⁵-fold diluted OD_{600nm} = 1 suspension onto the stigmata. Initial CFU densities of the inoculum were determined by mixing 10 μ L of the diluted suspension with 200 μ L PBS and plating 42 μ L on TSB agar plates. The plates were incubated at 26°C and CFU were counted after incubation. The inoculum was in the range of 20 to < 70 colonies.

For pre-inoculation of flowers with PW, a bacterial suspension of the strain with an OD_{600nm} = 1 was 10⁻³-fold diluted and 1.5 mL reaction tubes were filled up to the rim with the suspension. Stigmata of freshly opened flowers from GD trees grown in the greenhouse were inoculated by carefully dipping the stigmata into the suspension. On these flowers 2 μ L of a 10⁻⁴-fold diluted bacterial suspension of the selected *E. amylovora* strains were pipetted onto the pre-inoculated stigmata. For CFU determination of the inoculum, the 10⁻⁴-fold diluted bacterial suspensions were log diluted to a 10⁻⁵-fold dilution. Incubation and CFU determination was performed as described above. For assessing the microorganisms colonizing the orchard-P23 flowers, balloon flowers, and greenhouse flowers, in addition to the inoculated strains, dilutions of the reisolated bacterial suspensions were also plated on TSB agar plates supplemented with rifampicin (selection for fungi) and KB agar plates containing the fungicide cycloheximide (selection for bacteria) (Fig. S3).

DNA extraction. Bacterial DNA from apple flowers was extracted by transferring 0.5 mL of the 1 mL flower resuspension to sterile 1.5 mL reaction tubes followed by centrifugation for 1 min at 12.100 \times g in an Eppendorf mini centrifuge. The pellet was stored at -20°C until DNA was extracted using the BioSprint 96 DNA Plant kit (Qiagen) as described previously (39).

Real-time PCR to determine bacterial titer in flowers. Real-time PCRs were performed according to Pirc et al. (40) in a LightCycler 480 (Roche) using the TaqMan Universal master mix (Applied Biosystems) and the target gene *amsC*. The final reaction volume (10 μ L) contained 0.9 μ L of 10 μ M primer Ams116F (5'-TCCCACATACTGTGAATCATCCA-3'), 0.9 μ L of 10 μ M primer Ams189R (5'-GGGTA TTTGCGCTAATTTTATTCG-3'), 0.2 μ L of 10 μ M Ams141T (5'-FAM-CCA GAA TCT GGC CCG CGT ATA CCG-TAMRA-3'), 1 μ L ddH₂O, 5 μ L of 2 \times TaqMan universal master mix (Applied Biosystems), and 2 μ L of template DNA. All PCRs were conducted in triplicate and negative controls were included. The baseline was set automatically, cycling conditions were: 2 min at 50°C, 10 min at 95°C, 40 cycles of 15 s at 95°C and 1 min at 60°C. A crossing point (Cp) value above 38 would have been considered negative.

SUPPLEMENTAL MATERIAL

Supplemental material is available online only.

SUPPLEMENTAL FILE 1, PDF file, 1.5 MB.

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