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Measuring body composition and energy expenditure using (doubly) labelled water

Explanation of the principles and practical application

Sylvain Lerch
Agroscope, Ruminant Research Unit

13-14.09.2019, ISEP Course on Indirect Calorimetry, Belo Horizonte (BRA)

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Plan

Use of labelled water techniques in animal sciences

1. Labelled water technique for body composition measurement
 - Principles and applications
3. Doubly labelled water technique for energy expenditure measurement
 - Principles
4. Labelled water administration, sampling and analyses
 - Technical issues

Conclusions

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Interest in body composition and energy expenditure measurements in farm animals

Physiology **Animal Health** **Nutrition**

Understand body reserves management / allocation toward physiological functions Describe body reserves dynamics and energy expenditure across production cycle

Define animal requirements
⇒ Built-up / renovate feeding systems

Define carcass and products quality


Energy expenditure
productive and non-productive parts

Body composition
chemical: lipid, protein, ash, energy
tissular: adipose, muscles, bones, organs

Select most feed efficient individuals

Products quality **Genetics**

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Interest in body composition and energy expenditure measurements in farm animals

Gold-standard techniques

- **Body composition:** slaughter and dissection/grinding + analyses ⇒ destructive
- **Energy expenditure:** (in)direct calorimetry ⇒ confined chamber


↓

Need for *in vivo* techniques allowing

- Repeated measurements on a same individual
- Applicable in « real-life » rearing conditions / Reduced invasivity reduce physiological and behavioural perturbations

⇒ **Dosing animal with (doubly) labelled water and follow tracers elimination kinetics as a reliable technique?**

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Different types of labelled water for diverse applications

Singly labelled water:

$$\begin{array}{c} 18 \\ \text{H}-\text{O}-\text{H} \\ \text{H}_2^{18}\text{O} \\ \text{BG enrich.: 2000 ppm} \end{array}$$

$$\begin{array}{c} \text{D}-\text{O}-\text{D} \\ \text{}^2\text{H}_2\text{O or D}_2\text{O} \\ \text{BG enrich.: 150 ppm} \end{array}$$

$$\begin{array}{c} \text{T}-\text{O}-\text{T} \\ \text{}^3\text{H}_2\text{O or T}_2\text{O} \\ \text{Tritium water} \\ \text{radioactive} \end{array}$$

Applications

Labelled water dilution space ⇒ **Body composition**


+ Labelled water clearance ⇒ **Drinking water**

Doubly labelled water:

$$\begin{array}{c} 18\text{O} \\ \text{D}-\text{O}-\text{D} \end{array}$$

Differences in D and ^{18}O elim. rates ⇒ **Energy expenditure**

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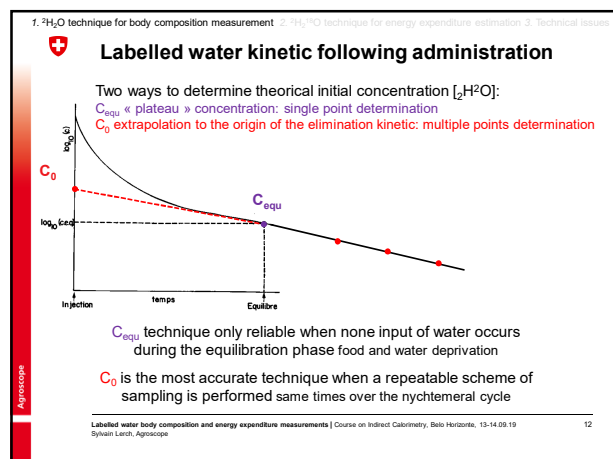
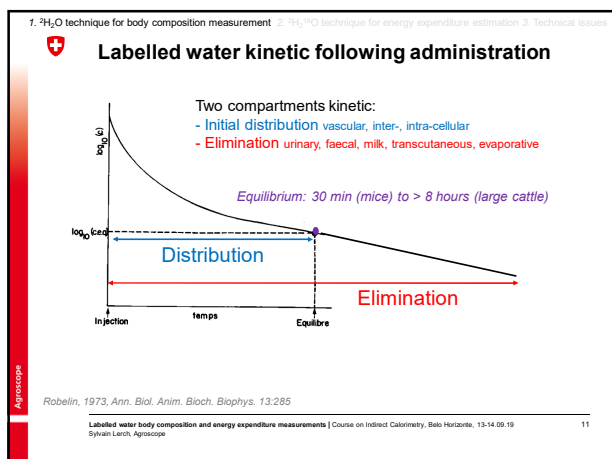
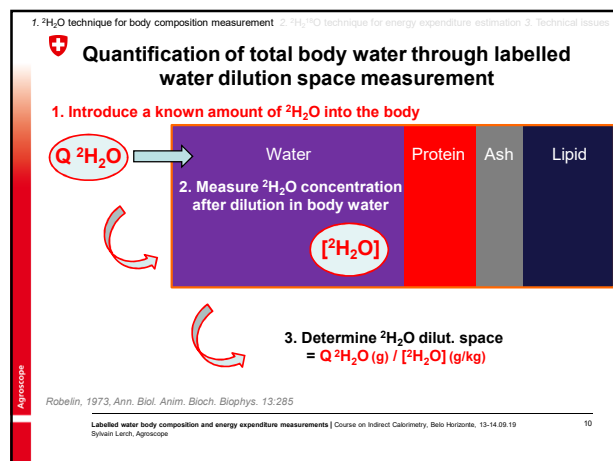
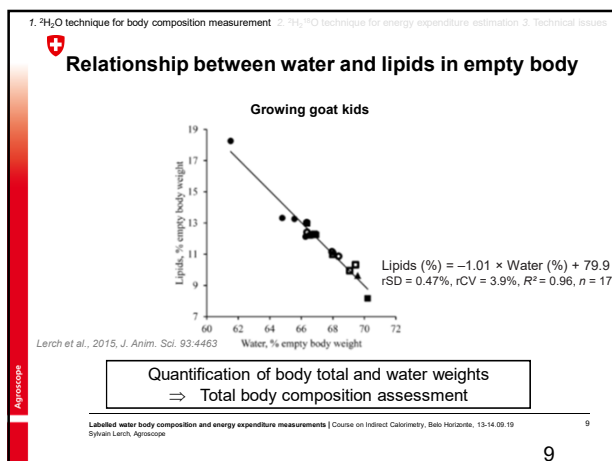
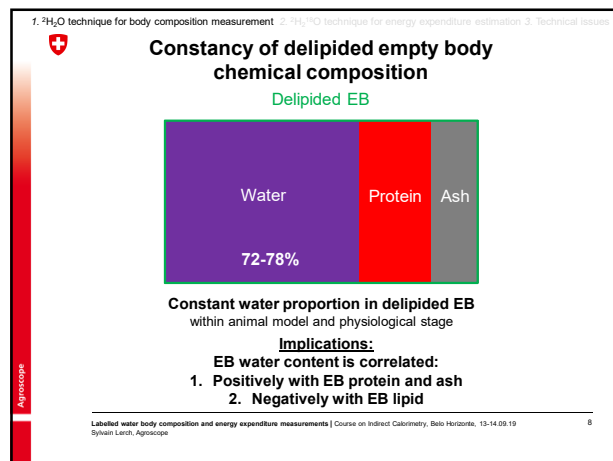
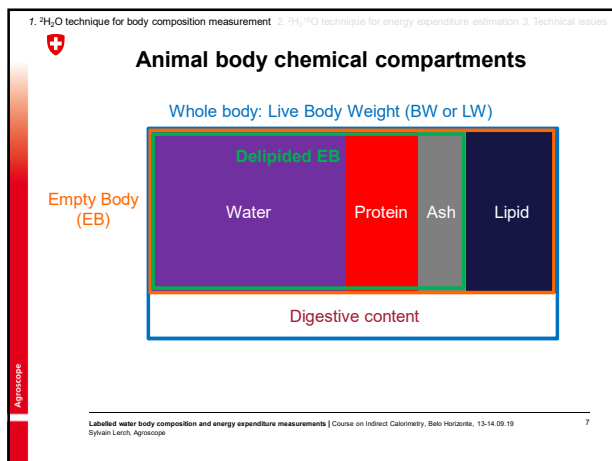
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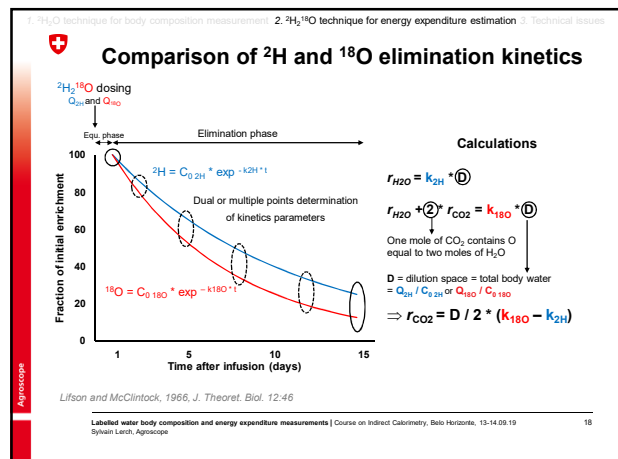
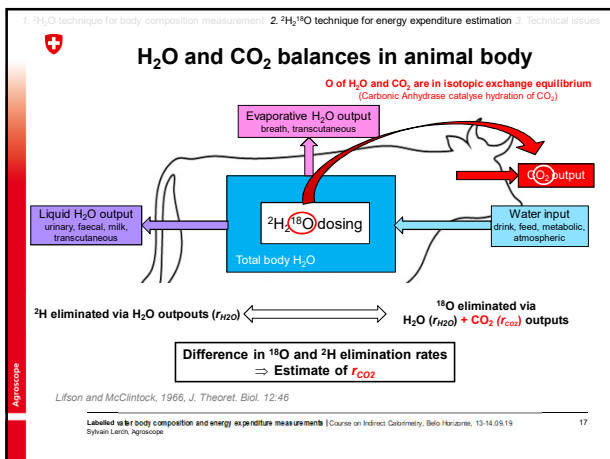
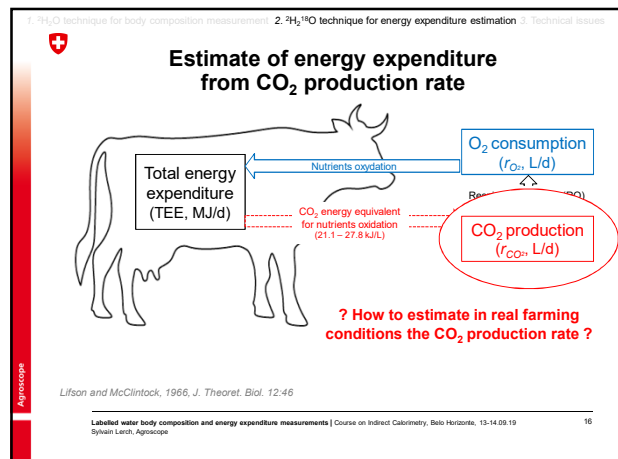
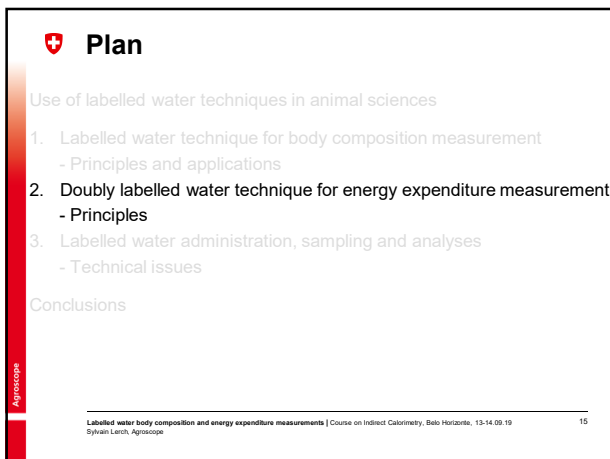
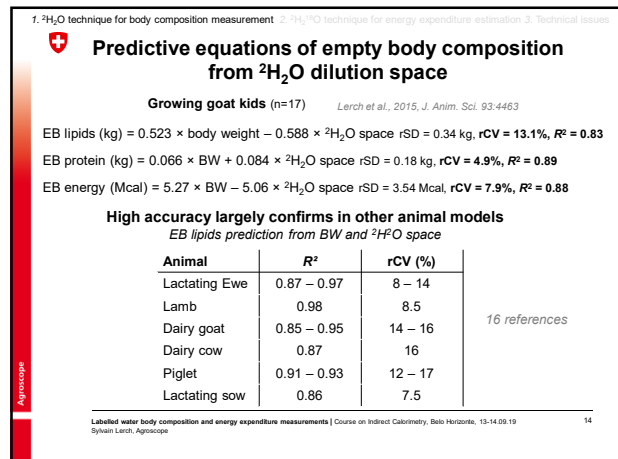
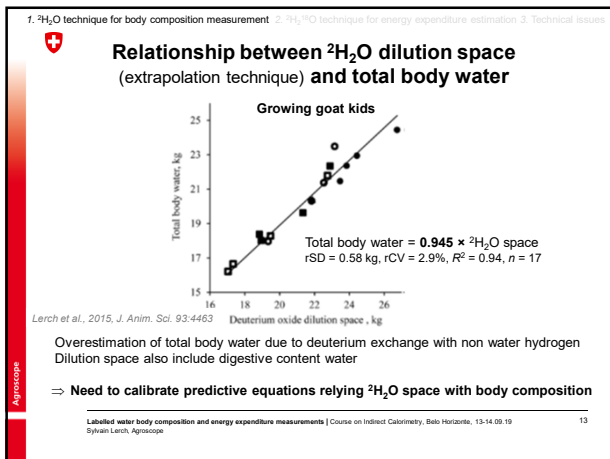
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Overview of the use of (doubly) labelled water techniques in animal sciences

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How to administrate and where to follow the kinetic of labelled water ?

Route of administration of the dose

Oral ingestion
Intramuscular injection
Intraperitoneal inj.
Intravenous injection

Risk of labelled water losses
Delay and variability in equilibration time
Risk of ^2H sequ. in dig.
Stress and invasivity

Compartments in which the elimination kinetic is follow

Blood => central compartment, homogeneous, could induce stress esp. for multiple sampling
Urine => elimination compartment, largely validate, needs the second collection urine
Saliva => reliable compared to blood (human), needs validation in farm animals!
(Milk) => Non-invasive alternative for lactating mammals

Speakman, 2001, *Bod. Comp. Anal.*:56

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Water extraction from samples

Reference method: water distillation / sublimation

From a « warm » to a « cold » point under vacuum

- Highly accurate and reproducible
- Specific « home made » equip.
- Time consuming 10-20 samp. / day

Robelin, 1973, *Biol. Anim. Bioch. Biophys.* 13:285
IPHC, CNRS Strasbourg (FR)
Cryo-distillation line, 2019

Alternative method: centrifuge tubes

Use of centrifuge tubes for deproteinization of blood plasma

- High throughput and convenient
- Uncompleted water extraction + isotope exchange => potential bias

Chery et al., 2015, *Rap. Comm. Mass Spectr.* 29:562

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Quantification of ^2H and ^{18}O enrichments

Infrared spectroscopy
Isotope-ratio mass spectrometry (IRMS)
Laser spectroscopy

1950
Calibration vs gradient enriched standards curve
Moderate accuracy and poor sensitivity
=> Need high enrichment level: high dosing

2000
Online reduction in TC-EA for $^2\text{H}^1\text{H}$ / $^1\text{H}^2\text{H}$ or $\text{C}^{18}\text{O}/\text{C}^{16}\text{O}$ gases ratio determination in IRMS
High accuracy and sensitivity
=> Fit with lower enrichment level (/5 vs IR spectroscopy)

2010
Injection of the sample as a gas in a tube scan by a laser
Fast and cheap
On-field applications
Accuracy in the frame of DLW study? Risks of artefacts?

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Approximate prices for $^2\text{H}_2\text{O}$ and $^2\text{H}_2^{18}\text{O}$ technics (in €)

Model size	$^2\text{H}_2\text{O}$ - body composition		$^2\text{H}_2^{18}\text{O}$ - energy expenditure	
	Equ. point n=2	Mult. points n=6	Dual points n=3	Mult. points n=6
Poultry ≈ 1.5 kg	40 (0.3%)*	120 (0.1%)	110 (8%)	220 (4%)
Sheep-goat ≈ 65 kg	45 (10%)	125 (4%)	500 (80%)	600 (65%)
Pig ≈ 120 kg	50 (17%)	130 (7%)	800 (85%)	900 (77%)
Cattle ≈ 600 kg	80 (50%)	160 (87%)	3'700 (97%)	3'900 (95%)

*% of total cost due to the dose is reported in brackets
Considering: - a dosing at 0.1 g $^2\text{H}_2\text{O}$ /kg BW and 0.05 g H_2^{18}O /kg BW
- a price of 0.7 €/g $^2\text{H}_2\text{O}$ and 120 €/g H_2^{18}O
- an analytical cost of 20€ / point for $^2\text{H}^1\text{H}$ and 35 € for both $^2\text{H}^1\text{H}$ and $^{18}\text{O}/^{16}\text{O}$ ratios determ.

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Conclusions: Deuterated water dilution space for *in vivo* body composition measurement

More than 85 years of use, still in use with several analytical improvement

- Need rigorous experimental procedures (injection, sampling), and specific analytical equipment and skills
- Non negligible cost

=> When calibrated predictive equations are used: accuracy only equalled/overpassed by few imaging techniques
CT-scan, MRI, DXA, need complete anaesthesia + not suitable for large cattle

Direct comparison of several techniques ($^2\text{H}_2\text{O}$ dil. space, US, CT, DXA, 3D imaging, BCS, adipocyte size, impedancemetry...) currently ongoing in goats
Pires, De La Torre, Lerch et al.

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Alternative to indirect calorimetry, suitable for on-field study

Accuracy relative depending on corrections for isotope exchange/incorporation especially for ^2H during fat accretion in fast growing animal

Extremely expensive: until now only few applications in farm animal
< 10 published articles, reduced sized models in most of the cases

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